Ministry for Primary Industries
Manatū Ahu Matua

Animal Products Notice

Recognised Laboratories

15 November 2024

Issued under the Animal Products Act 1999

New Zealand Government

TITLE

Animal Products Notice: Recognised Laboratories

COMMENCEMENT

Parts 1 to 5 and Schedule 1 of this Animal Products Notice comes into force on 10 December 2024.

Schedule 2 comes into force on 1 January 2025.

REVOCATION

This Animal Products Notice revokes and replaces the Animal Products Notice: Recognised Laboratories, issued on 09 May 2022.

This Animal Products Notice revokes Schedule 1 on 31 December 2024.

ISSUING AUTHORITY

This Animal Products Notice is issued under section 167(1) for the purposes of section 77H of the Animal Products Act 1999 and under section 167(2) of the Animal Products Act 1999 to supplement Part 9 of the Animal Products Regulations 2021.

Dated at Wellington, 15 November 2024

[Signed]

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Director, Food Risk Management
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(acting under delegated authority of the Director-General)

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Contents

Page

Introduction **3**

Part 1: Preliminary **5**
1.1 Application 5
1.2 Definitions 5

Part 2: Recognition requirements **6**
2.1 Accreditation 6
2.2 Making documentation available 6
2.3 Non-compliance reporting 6
2.4 Annual reporting - live animal and germplasm testing 6
2.5 Annual reporting - bivalve molluscan shellfish 6

Part 3: General testing **8**
3.1 Acceptable test methods 8
3.2 Composite samples 8
3.3 Test reports for sub-contracted tests 8

Part 4: Bivalve molluscan shellfish testing **9**
4.1 Application 9
4.2 Phytoplankton testing qualification 9
4.3 Test methods 9
4.4 Sample receipt 9
4.5 Sample storage 10
4.6 Testing and reporting timeframes 10
4.7 Method performance 12
4.8 Reporting results 12

Part 5: Designated Inter-Laboratory Comparison Programme **15**
5.1 When to participate 15
5.2 Procedure for participation 15
5.3 Defect categories 15
5.4 Minor and major defect categories 15
5.5 Critical defect category 16
5.6 Resumption of designated ILCP testing 16

Schedule 1: Designated ILCP tests **17**

Schedule 2: Designated ILCP tests **19**

Introduction

This introduction is not part of the Animal Products Notice, but is intended to indicate its general effect.

Purpose

(1) The purpose of this notice is to supplement the Animal Products Regulations 2021 and set requirements for recognised laboratories.

Background

(1) The Animal Products Act 1999 sets the framework for the recognition of agencies.

(2) The Animal Products Regulations 2021 sets specific requirements for recognised agencies, with additional requirements for recognised laboratories.

Who should read this Animal Products Notice?

(1) This Animal Products Notice should be read by:

a) laboratories applying to be recognised agencies; and
b) recognised laboratories; and
c) persons conducting testing in a recognised laboratory or performing any specialist laboratory function or testing activity in connection with such laboratories; and
d) animal product exporters, processors and risk management programme operators who contract recognised laboratories to perform testing to satisfy their obligations under the Animal Products Act 1999.

Why is this important?

(1) A laboratory that fails to comply with the requirements of this Notice may not be granted, or may not retain, as applicable, recognition.

(2) A failure to comply with this Notice may be an offence under the Animal Products Act 1999.

Document History

Version Date	Section Changed	Change(s) Description
09 May 2022	-	New Notice
15 November 2024	1.2 2.4 2.5 3.1(1) Part 4 (new) Part 5 (renumbered) Schedule 1 Schedule 2	Added definitions Added definition Added BMS annual reporting (previously in BMS Notice) Added exception for modifying test methods Added a new part to cover testing requirements for BMS (previously in BMS Notice) Renumbered (from part 4 to 5), added references to Schedule 2, and corrected clause numbering To be revoked on 31 Dec 2024, renumbered tables New schedule for Designated ILCP testing, to take force on 1 Jan 2025

Other information

(1) Laboratories are also subject to relevant requirements including the following legislation:

a) Animal Products Act 1999;
b) Animal Products Regulations 2021.

Part 1: Preliminary

1.1 Application

(1) This Notice applies to a laboratory recognised under section 101 of the Animal Products Act 1999.

(2) This Notice does not apply to a laboratory recognised under section 102 of the Act.

1.2 Definitions

(1) In this Notice, unless the context otherwise requires:
Act means the Animal Products Act 1999

BMS means bivalve molluscan shellfish

BMS Notice means the Animal Products Notice: Regulated Control Scheme – Bivalve Molluscan Shellfish for Human Consumption

designated ILCP means the designated Inter-Laboratory Comparison Programme, which is a proficiency programme operated on behalf of the Ministry for Primary Industries

designated ILCP provider means the person contracted by the Ministry for Primary Industries to operate the designated ILCP

IANZ means International Accreditation New Zealand

ILCP means Inter-Laboratory Comparison Programme

ISO/IEC 17025 means NZS ISO/IEC 17025:2018 *General requirements for the competence of testing and calibration laboratories*

MPI means Ministry for Primary Industries

Regulations means the Animal Products Regulations 2021; and a reference to a specific Regulation is a reference to that regulation in those Regulations

(2) Any term defined in the Act, the Animal Products Regulations 2021, Animal Products Notice: Production, Supply and Processing, Animal Products (Regulated Control Scheme – Bivalve Molluscan Shellfish) Regulations 2006 or Animal Products Notice: Regulated Control Scheme - Bivalve Molluscan Shellfish for Human Consumption that is used in this Notice but not defined has the meaning given in the Act, Regulations, or that Notice.

Part 2: Recognition requirements

2.1 Accreditation

(1) For the purposes of Regulation 190(1)(b)(i), a recognised laboratory must be accredited to ISO/IEC 17025.

2.2 Making documentation available

(1) A recognised laboratory must provide employees performing tests and other relevant functions with access to:

a) the current version of the Act, and all relevant regulations, notices and directions made under it; and
b) ISO/IEC 17025 and any other documents or information related to the laboratory's accreditation; and
c) relevant parts of the laboratory's own systems, procedures, records, and databases.

2.3 Non-compliance reporting

(1) For the purposes of Regulation 206(2), a report to the Director-General about a non-compliance must include:

a) a description of what has occurred; and
b) the actions taken or to be taken to ensure integrity of test results; and
c) the actions taken or to be taken to address the root cause.

2.4 Annual reporting - live animal and germplasm testing

(1) A recognised laboratory that performs live animal or germplasm testing (testing done to support the export of live animals (not for human consumption) and germplasm, with or without an official assurance), must provide an annual report for all export testing carried out on live animals or germplasm for the period 1 July to 30 June, by 31 July of each calendar year.

(2) The annual reports must include:

a) the name of the species of animal that is tested; and
b) the number and type of tests performed; and
c) the results of those tests; and
d) results of ILCP testing, including full ILCP laboratory report; and
e) details of non-conformances identified in internal audits; and
f) any identified conflicts of interest and any subsequent mitigation strategies.

2.5 Annual reporting - bivalve molluscan shellfish

(1) A recognised laboratory that has the tests listed in 4.3, plus heavy metal testing of BMS flesh, listed in their recognition scope, must provide an annual report for all testing performed for the period 1 January to 31 December, by 31 January of the following calendar year.

(2) The annual report must include all of the following:

a) the number of samples tested, including:

i) the nature of the sample; and

ii) the species of BMS, where applicable; and
iii) the method used; and

b) the number of samples received in a non-conforming state, the nature of the non-conformance, and whether the sample was tested or disposed of; and
c) a summary of any inter-laboratory comparison programme (ILCP) activity including reports, results and any corrective actions, for the previous year; and
d) the proposed ILCP schedule, for biotoxin and phytoplankton testing only, indicating the testing rounds of participation for each assay and matrix for the coming year; and
e) any major non-conformances detected by IANZ or assessments initiated by the Director-General.

Part 3: General testing

3.1 Acceptable test methods

(1) If a test is required under the Act to be done by a specified test method, a recognised laboratory must use the specified test method, without modification, except when testing for biotoxin or phytoplankton (4.3 (1) and (2)), where modifications may be made.

(2) If no test method is specified, a recognised laboratory must use a test method suitable for the relevant sample matrix.

3.2 Composite samples

(1) A recognised laboratory may combine samples into a composite sample only:

a) if requested by the client; and
b) for the purpose of microbiological or chemical testing; and
c) where discrete testing of a specified quantity of representative samples is not required under the Act.

(2) A recognised laboratory may combine samples of animal material, or of animal product, or environmental samples into a composite sample.

(3) A recognised laboratory must have records that link:

a) the information supplied with the original samples to the composite sample; and
b) the information about the composite sample to the original samples.

(4) A recognised laboratory that combines samples into a composite sample for microbiological or chemical testing must:

a) have defined procedures for combining samples to form a composite sample and the aliquot of the composite sample selected; and
b) determine and record that the use of a composite sample is appropriate for the test method, including:

i) the maximum sample size that is suitable for the test; and
ii) the maximum number of individual samples that can be included in a composite sample; and
iii) the level of sensitivity that will be achieved by testing a composite sample compared to discrete sample testing.

(5) Microbiological testing of a composite sample may be used to determine presence or absence of particular pathogens but may only be used for enumeration of a pathogen if the laboratory has considered the limitations of composite testing for enumeration, such as reduced sensitivity.

(6) Any report providing test results for a composite sample must clearly identify the individual samples used to create the composite sample.

3.3 Test reports for sub-contracted tests

(1) Any report providing test results where the testing has been subcontracted to another recognised laboratory (see Regulation 204) or to an overseas laboratory (see Regulation 205), must contain information that enables the following to be identified:

a) the subcontracted laboratory; and
b) the individual or individuals who authorised the results.

Part 4: Bivalve molluscan shellfish testing

4.1 Application

(1) Part 4 applies to a recognised laboratory that has the tests listed in 4.3, plus heavy metal testing of BMS flesh, listed in their recognition scope.

4.2 Phytoplankton testing qualification

(1) A recognised laboratory that conducts analyses of seawater samples for toxigenic phytoplankton must employ a phytoplankton taxonomist who:

a) has attended an Intergovernmental Oceanographic Commission Advanced Course in Phytoplankton Taxonomy; or
b) successfully participates annually in the International Phytoplankton Intercomparison (IPI) proficiency test for marine phytoplankton.

4.3 Test methods

(1) For testing BMS flesh for biotoxins, a recognised laboratory must use the following methods:

a) Paralytic Shellfish Poison (PSP) - J. AOAC (2020) Int 103(2): 533-562; and
b) Amnesic Shellfish Poison (ASP) screen test - J. AOAC Int 2005 88(3): 761-772; and
c) Amnesic Shellfish Poison (ASP) confirmation test – J. AOAC Int 1995 78(2): 543-554; and
d) Diarrhetic Shellfish Poison (DSP) - J. AOAC Int 2005 88(3): 761-772; and
e) Neurotoxic Shellfish Poison (NSP) screen test - J. AOAC Int 2005 88(3): 761-772; and
f) Neurotoxic Shellfish Poison (NSP) confirmation test - J. AOAC. Int. 2012 95(4): 1097-1105; and
g) Azaspiracid Shellfish Poison (AZP) - J. AOAC Int 2005 88(3): 761-772.

(2) For analysing seawater samples for toxigenic phytoplankton, a recognised laboratory must use the following methods:

a) UNESCO Phytoplankton Manual (1978) and IOC Manual & Guides 55 (2010); and
b) for DNA Probe (*Pseudo-nitzschia* speciation), Scholin C.A. et al 1996. Phycologia 35: 190-197.

(3) For testing *Escherichia coli* in BMS flesh, a recognised laboratory must use the MPI method: Enumeration of *Escherichia coli* in Molluscan Bivalve Shellfish.

(4) For testing seawater for faecal coliforms, a recognised laboratory must use the 4th edition of the American Public Health Association's Recommended Procedures for the Examination of Sea Water and Shellfish.

(5) For tests from clause (1), a recognised laboratory must propose any changes to the following agreed criteria, in writing:

a) any Toxicity Equivalence Factors used in biotoxin calculations; and
b) any levels of biotoxin in a screen test that trigger use of a confirmation test.

4.4 Sample receipt

(1) A recognised laboratory must check the following on receipt of a sample:

a) the person who took the sample holds a current certificate of competency as a sample taker issued by an Animal Product Officer under the BMS Notice; and
b) the sample provided is suitable for the particular test required; and

c) for biotoxin, phytoplankton and microbiological samples, that the sample was received in accordance with Table 1; and
d) for microbiological samples, that the sample temperature is less than 10°C, unless:

i) sampling occurred on the same day; and
ii) if placed in a chilled container, the sample has not had adequate time to reach temperatures lower than 10°C.

(2) If any of the requirements of clause (1) are not met, or if the recognised laboratory considers the sample may not be suitable for testing, the recognised laboratory must:

a) decide whether to analyse the sample or to seek direction from the APO responsible for the growing area; and
b) notify the sampler and the APO within 1 working day of sample receipt; and
c) if the decision is taken that a replacement sample is necessary, analyse as a priority any replacement sample; and
d) if an extension to reporting timeframes in Table 1: Testing and reporting timeframes is needed, seek approval in accordance with 4.6 (4), (5) and (6).

(3) If any of the requirements of clause (1) are not met, records must be kept of:

a) which requirements were not met for each sample; and
b) whether the recognised laboratory decided to analyse the sample or the direction received from the APO; and
c) all notifications to samplers and APOs under clause (2).

4.5 Sample storage

(1) A recognised laboratory must maintain biotoxin and microbiological samples at a temperature of less than 4°C until analysis is started.

(2) If a sample is required for an official investigation, it must be stored until the recognised laboratory has been notified in writing by MPI that the sample may be discarded.

4.6 Testing and reporting timeframes

(1) A recognised laboratory must test and report biotoxin, phytoplankton and microbiological samples in accordance with Table 1: Testing and reporting timeframes.

Table 1: Testing and reporting timeframes

Sample	Test	Start time	Completion time
BMS flesh (live samples only)	Biotoxins	Test must start within 48 hours from sample collection, otherwise follow 4.4(2).	Testing and reporting must be completed within 2 working days after sample receipt. Samples received on Friday afternoon may be reported on the next Wednesday. Reporting to an MPI database is required, as per 4.8(1).
	Escherichia coli	Test must start within 24 hours from sample collection, otherwise follow 4.4(2) and 4.6(2).	Testing and reporting must be completed within 3 working days after sample receipt.
Seawater	Faecal coliforms		Testing and reporting must be completed within 5 working days after sample receipt.
	Phytoplankton	Test must start within 48 hours from sample collection, otherwise follow 4.4(2).	Testing and reporting must be completed within 24 hours after sample receipt. Reporting to an MPI database is required, as per 4.8(1).

(2) Testing for *Escherichia coli* in live BMS flesh and faecal coliforms in seawater may start within 48 hours from sample collection if:

a) transport delays have occurred; and
b) the delay is documented; and
c) the temperature requirements in clause 4.4 (1) h) are met.

(3) A shellfish specialist may give written approval for a recognised laboratory to extend a reporting timeframe on a routine basis on the grounds that:

a) the test method routinely requires longer periods; or
b) complex confirmatory procedures make a definitive time period difficult to estimate.

(4) A shellfish specialist may give written approval for a recognised laboratory to extend a reporting timeframe for a particular sample or group of samples on the grounds that:

a) a technical failure has been encountered in the recognised laboratory; or
b) there were transport difficulties with the sample; or
c) the recognised laboratory is closed, for example for a public holiday.

(5) Any request for an extension to the reporting timeframe must be raised in writing by a recognised laboratory and must describe:

a) the test to be performed; and
b) the justification for extension; and
c) the revised suggested reporting time; and
d) any corrective actions that will be taken to resolve any technical failure.

(6) Records must be kept of:

a) decisions to extend reporting timeframes made under (2), (3) and (4); and
b) written approval given under (3) and (4), for as long as it is relied on, but for a minimum of 4 years from the date given.

4.7 Method performance

(1) A recognised laboratory must ensure that samples in a batch are re-analysed where:

a) batch control values are outside the limits or requirements of the method performance standards; and
b) the laboratory considers this may affect the results.

(2) If there is an unidentified test response for biotoxins, a laboratory must:

a) notify the Director-General within 24 hours; and
b) investigate the response, which may include:

i) testing samples in duplicate;
ii) repeat testing of a sample, as long as the remainder of the sample is sufficient for that process;
iii) sending samples for testing to another laboratory, if the remainder of the sample is sufficient for that process; and
iv) independent confirmation testing, at the laboratory or at another appropriate laboratory; and

c) if possible, identify the unknown compound.

(3) The laboratory must provide the Director-General with a report of all unidentified test response findings once the investigation (from (2)(b)) is complete.

4.8 Reporting results

(1) If required by Table 1, a recognised laboratory must report analytical results and sample information into an MPI database within 24 hours after each result is confirmed.

(2) Biotoxin results must be reported in the units described in Table 3: Maximum permissible levels for marine biotoxins in BMS.

(3) Biotoxin results in the test reports must clearly show:

a) which toxins are required to be summed for each toxin group; and
b) the total level obtained for each of those toxin groups.

(4) Biotoxin results obtained below the limit of detection of the test method must be reported as 'Not Detected'.

(5) If any of the following results are obtained, they must be reported verbally (or by a method acceptable to a shellfish specialist) by the laboratory to an APO responsible for the relevant growing area (or a nominated representative) within 1 hour after confirmation of the result, and be confirmed in writing within 24 hours:

a) *Escherichia coli* levels in BMS greater than 230 MPN per 100 grams; and
b) faecal coliform levels in seawater greater than 14 MPN per 100 ml; and
c) levels of toxigenic phytoplankton greater than the trigger levels stated in Table 2: Toxigenic phytoplankton action levels in seawater; and
d) levels of biotoxins in BMS greater than the maximum permissible levels shown in Table 3: Maximum permissible levels for marine biotoxins in BMS.

Table 2: Toxigenic phytoplankton action levels in seawater

Phytoplankton species	Marine biotoxin group	Level in composite sample to trigger BMS flesh testing.[1] Cells per litre (c/L)
Alexandrium minutum	PSP	100
Alexandrium ostenfeldii	PSP	100
Alexandrium catenella	PSP	100
Alexandrium pacificum	PSP	100
Alexandrium tamarense	PSP	100
Gymnodinium catenatum	PSP	100
[2]*Pseudo-nitzschia australis* [2]*Pseudo-nitzschia pungens* [2]*Pseudo-nitzschia multiseries* [2]*Pseudo-nitzschia multistriata*	ASP	100,000
[2]*Pseudo-nitzschia fraudulenta* [2]*Pseudo-nitzschia delicatissima* [2]*Pseudo-nitzschia pseudodelicatissima*	ASP	1,000,000
[3]*Karenia brevis*	NSP	1,000
Karenia brevisculata	NSP	10,000
[4]*Karenia/Karlodinium/Gymnodinium* Group	NSP	250,000
Dinophysis acuta	DSP	500
Dinophysis acuminata	DSP	1,000
Prorocentrum lima	DSP	500
Azadinium Group	AZP	30,000

[1] If the trigger level is exceeded, a BMS sample must be taken within 24 hours of notification of the trigger level and submitted for analysis for the relevant toxin.

[2] For *Pseudo-nitzschia* species, if the 100,000 c/L level is exceeded a DNA probe or BMS analysis for domoic acid must be performed.

BMS analysis for domoic acid must be performed if the combined percentage result from a DNA probe for the 4 most toxic species (*P. australis, [2]P. multistriata P. multiseries* and *P. pungens*) exceeds 100,000 if applied to the original cell count (preserved seawater sample).

BMS analysis for domoic acid must be performed if the combined percentage result from a DNA probe for the 3 less toxic species exceeds 1,000,000 c/L if applied to the original cell count (preserved seawater sample).

If DNA probe speciation is not performed, the default trigger level of 100,000 c/L applies.

[3] *Karenia brevis* has not been isolated in New Zealand to date.

[4] The *Karenia/Karlodinium/Gymnodinium* group includes *Karenia bidigitata, Karenia mikimotoi, Karenia papilionacea, Karenia selliformis, Karlodinium micrum* and *Gymnodinium impudicum.*

Table 3: Maximum permissible levels for marine biotoxins in BMS

Toxin group	Amount that must not be exceeded in the edible portion
Paralytic Shellfish Poison (PSP)	0.8 mg saxitoxin dihydrochloride equivalent per kg
Amnesic Shellfish Poison (ASP)	20 mg domoic acid per kg
Neurotoxic Shellfish Poison (NSP)	0.8 mg brevetoxin-2 equivalent per kg
Diarrhetic Shellfish Poison (DSP)	0.16 mg of okadaic acid[1] equivalent per kg
Azaspiracid Shellfish Poison (AZP)	0.16 mg of azaspiracid-1[2] equivalent per kg

[1] Okadaic acid includes okadaic acid (OA), dinophysistoxins (DTX1 and DTX2). For okadaic acid and dinophysistoxins a hydrolysis step is required in order to detect the presence of esterified okadaic acid group toxins.

[2] Azaspiracid includes AZA1, AZA2 and AZA3.

Part 5: Designated Inter-Laboratory Comparison Programme

5.1 When to participate

(1) A recognised laboratory that has any of the tests set out in Schedule 1: Designated ILCP Tests or Schedule 2: Designated ILCP Tests in its scope of recognition must participate in the designated ILCP (see Regulation 200(2)).

5.2 Procedure for participation

(1) Every participating laboratory must advise the designated ILCP provider of:

a) the designated ILCP tests that are in its scope of recognition when first recognised; and
b) any subsequent addition to, or removal from designated ILCP tests in its scope of recognition; and
c) any suspension of all or part of its recognition where that suspension includes designated ILCP tests.

(2) For each designated ILCP test that a participating laboratory has in its scope of recognition, the laboratory must perform at least the number of rounds of that test specified in Schedule 1: Designated ILCP Tests or Schedule 2: Designated ILCP Tests.

(3) On receipt of a set of proficiency samples from the designated ILCP provider for a round of tests, the laboratory must begin testing:

a) within 48 hours of receipt or the round opening, in the case of microbiological tests; and
b) as soon as practicable after receipt or the round opening, in the case of chemical tests.

(4) The laboratory must return the results of tests to the designated ILCP provider by the date specified by the designated ILCP provider as the date on which the round closes.

(5) The laboratory is liable for the cost of all samples (including samples provided for re-tests) provided by the designated ILCP provider.

5.3 Defect categories

(1) The designated ILCP provider may categorise performance that is below expectations as one of the following:

a) minor defect category; or
b) major defect category; or
c) critical defect category.

5.4 Minor and major defect categories

(1) If the designated ILCP provider notifies a participating laboratory of a minor or major defect category, the laboratory must:

a) request a re-test sample within 2 days after receipt of the notification; and
b) complete testing of the re-test samples within 5 days after receipt of the samples and return the results to the designated ILCP provider; and
c) carry out and document an investigation into the cause; and
d) take any necessary corrective action.

5.5 Critical defect category

(1) If the designated ILCP provider notifies a participating laboratory of a critical defect category, the laboratory must:

a) continue participation in the minimum number of rounds specified in the Tables in Schedule 1: Designated ILCP Tests or Schedule 2: Designated ILCP Tests for the tests it does; and
b) demonstrate its capability by:

i) identifying and undertaking appropriate corrective actions to regain testing proficiency; and
ii) undertaking any repeat testing required by the designated ILCP provider and report back to the designated ILCP provider on results obtained.

5.6 Resumption of designated ILCP testing

(1) Sub-clause 5.6(2) applies when a participating laboratory recommences performing designated ILCP tests after a laboratory has been closed temporarily or all or part of its recognition is suspended and that suspension includes designated ILCP tests.

(2) For the purpose of calculating how many rounds of designated ILCP tests have been done by the participating laboratory in a year, the round immediately preceding the closure or suspension of all or part of its recognition must be treated as its most recent designated ILCP round.

Schedule 1: Designated ILCP tests

Note: Schedule 1 will be revoked from 31 December 2024.

Table 4: Potable/Suitable Water Microbiology Comparative Programme (meat, poultry and fish)

		Designated ILCP	
Numerical reference	**Test name**	**Designated ILCP test name**	**Minimum number of rounds per year**
1.3	Colony count 22°C	Standard plate count SPC22	6
1.1.1	Total coliforms (coliform bacteria), *Escherichia coli*	Total coliforms, *E. coli*	6
11.1.2			
1.2	Faecal coliforms	Faecal coliforms	6
11.1.1			
1.6.1	*Clostridium perfringens* (including spores when available but no less than 2 rounds)	*Clostridium perfringens*	6
11.6.1	Faecal coliforms (growing water)	Faecal coliforms	4

Table 5: Meat and Poultry Microbiology Comparative Programme

		Designated ILCP	
Numerical reference	**Test name**	**Designated ILCP test name**	**Minimum number of rounds per year**
2.1.1	Aerobic Plate Count (APC)	APC 30 (SPC)	11
2.1.2	APC spread plate		
2.1.3	APC Petrifilm		
2.1.4	APC spiral plater		
2.2.1	*Escherichia coli*, direct plate or Petrifilm	*E. coli*	11
2.2.2	*Escherichia coli*, Petrifilm		
2.3	*Staphylococcus aureus*	*Staphylococcus aureus*	2
2.8 & 2.8.1	*Clostridium perfringens*	*Clostridium perfringens*	2
2.9	Enterobacteriaceae	Enterobacteriaceae	2
2.10	Faecal coliforms	Faecal coliforms	2

Table 6: Fish Microbiology Comparative Programme

		Designated ILCP	
Numerical reference	**Test name**	**Designated ILCP test name**	**Minimum number of rounds per year**
11.5.3	Total Plate Count (TPC) or Aerobic Plate Count (APC)	APC (SPC)	4
11.6.7	APC, TPC		
11.8.6	APC		
11.5.10	*Escherichia coli*	*E. coli*	4
11.6.2			
11.8.1			
11.5.4	*Staphylococcus aureus*	*Staphylococcus aureus*	4

Table 7: Pathogen Microbiology Comparative Programme (meat, poultry and fish)

		Designated ILCP	
Numerical reference	**Test name**	**Designated ILCP test name**	**Minimum number of rounds per year**
2.4.1	*Salmonella*	*Salmonella*	2 rounds per year for each *Salmonella* method the laboratory is recognised for
2.4.2	*Salmonella*		
2.4.3	*Salmonella*		
11.5.11	*Salmonella*		
11.6.3	*Salmonella*		
11.8.2	*Salmonella*		
2.6	*Listeria monocytogenes*	*Listeria monocytogenes* presence/absence and enumeration	2 rounds per year for each *Listeria monocytogenes* method the laboratory is recognised for
11.8.5	*Listeria monocytogenes*		
22.1	*Campylobacter*	*Campylobacter* enumeration	2
23.1	*Escherichia coli* O157:H7	*Escherichia coli* O157:H7 and Top 6 nSTECs	2
23.3	Top 7 Shiga Toxin-producing *Escherichia coli*		
11.5.12	*Vibrio cholerae*	*Vibrio* species	2 rounds per year for each *Vibrio* species and matrix the laboratory is recognised for
11.6.9	*Vibrio cholerae*		
11.5.6	*Vibrio parahaemolyticus*		
11.6.4	*Vibrio parahaemolyticus*		

Table 8: Potable/Suitable Water Chemistry Comparative Programme; all markets – surveillance of potable water in meat & game export premises

		Designated ILCP	
Numerical reference	**Test name**	**Designated ILCP test name**	**Minimum number of rounds per year**
5.02	Conductivity	Conductivity	2
5.03	pH (hydrogen ion concentration)	pH	2
5.04	Turbidity	Turbidity	2
5.10	Ammoniacal nitrogen (ammonium)	Ammoniacal nitrogen (ammonium)	2
5.13	Nitrate	Nitrate	2
5.14	Nitrite	Nitrite	2

Schedule 2: Designated ILCP tests

Note: Schedule 2 comes into force on 1 January 2025.

Table 9: Potable/Suitable Water Microbiology (meat, poultry and fish)

		Designated ILCP	
Numerical reference	**Test name**	**Designated ILCP test name**	**Minimum number of rounds per year**
1.3	Colony count 22°C	Standard plate count SPC22	6
1.1.1	Total coliforms (coliform bacteria), *Escherichia coli*	Total coliforms, *E. coli*	6
1.2	Faecal coliforms	Faecal coliforms	6
1.6.1	*Clostridium perfringens* (including spores when available)	*Clostridium perfringens*	6
11.6.1	Faecal coliforms (growing water)	Faecal coliforms	4

Table 10: Meat, Poultry, Fish and Dairy Microbiology

		Designated ILCP	
Numerical reference	**Test name**	**Designated ILCP test name**	**Minimum number of rounds per year**
2.1.1	Aerobic Plate Count (APC)	APC 30 (SPC)	11
2.1.2	APC spread plate		
2.1.3	APC Petrifilm		
2.1.4	APC spiral plater		
2.2.1	*Escherichia coli*, direct plate or Petrifilm	*E. coli*	11
2.2.2	*Escherichia coli*, Petrifilm		
2.9	Enterobacteriaceae	Enterobacteriaceae	2
11.5.10	*Escherichia coli*	*E. coli*	4
11.6.2			
11.8.1			
31.2	*Bacillus cereus*	*Bacillus cereus*	2

Table 11: Pathogen Microbiology (meat, poultry, fish and dairy)

		Designated ILCP	
Numerical reference	**Test name**	**Designated ILCP test name**	**Minimum number of rounds per year**
2.4.1	*Salmonella*	*Salmonella*	2 rounds per year for each *Salmonella* method the laboratory is recognised for
2.4.2	*Salmonella*		
2.4.3	*Salmonella*		
11.5.11	*Salmonella*		
11.6.3	*Salmonella*		
11.8.2	*Salmonella*		
31.12	*Salmonella*		
2.6	*Listeria monocytogenes*	*Listeria monocytogenes* presence/absence and enumeration	2 rounds per year for each *Listeria monocytogenes* method the laboratory is recognised for
11.8.5	*Listeria monocytogenes*		
31.10	*Listeria monocytogenes*		
22.1	*Campylobacter*	*Campylobacter* enumeration	2
23.1	*Escherichia coli* O157:H7	*Escherichia coli* O157:H7 and Top 6 nSTECs	2
23.3	Top 7 Shiga Toxin-producing *Escherichia coli*		