



Guidance Document

Semen and Embryos from Equids

EQUIGERM.SPE

15 March 2023

Title

Guidance Document: Semen and Embryos from Equids

About this document

This guidance document contains information about acceptable ways of ensuring compliance with the requirements in the *Import Health Standard (IHS): Semen and Embryos from Equids*.

Any guidance on how to comply with the applicable requirements may not be the only way to achieve compliance. Stakeholders are encouraged to discuss departures from the approaches outlined in this guidance document with the Ministry for Primary Industries (MPI) to avoid expending resources on the development of alternative approaches which may later be considered unsuitable.

The term “must” is not typically used in guidance. In this particular document if the term “must” is used, it is used in the context of quoting or paraphrasing the requirements set out in the related *IHS: Semen and Embryos from Equids*.

Related Requirements

Import Health Standard: Semen and Embryos from Equids

Document history

Refer to Appendix 1.

Contact Details

For further information and questions about this guidance document, please contact:

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Disclaimer

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1 Purpose

- (1) This guidance document has been issued to accompany the *IHS: Semen and Embryos from Equids*. This guidance document should be read in conjunction with that IHS.
- (2) This document includes:
 - a) Countries with MPI-approved exporting systems to export equine semen and embryos to New Zealand.
 - b) Model veterinary certificates for semen and embryos.
 - c) Links to negotiated country specific veterinary certificates.

2 Background

- (1) The *IHS: Semen and Embryos from Equids*, which this guidance document accompanies, contains generic import requirements. These are the rules to manage the biosecurity risk of importing equine semen and embryos from the countries specified in the IHS (Australia, Canada, EU member countries, Norway, Switzerland, and the USA) and in doing so meet New Zealand's appropriate level of protection. This guidance document contains model veterinary certificates for trade in equine semen and embryos. This veterinary certificate represents what will be certified prior to exporting consignments of equine semen and embryos from the country specified.
- (2) General information about importing semen and embryos can be found here: <http://mpi.govt.nz/importing/live-animals/semen-and-embryos/>.

3 Definitions

- (1) Refer to Schedule 2 of the *IHS: Semen and Embryos from Equids*.

4 Importer Responsibilities

- (1) The costs to MPI in performing functions relating to the importation of equine semen and embryos will be recovered in accordance with the Biosecurity Act 1993 (the Act) and any regulations made under that Act. All costs involved with documentation, transport, storage and obtaining a biosecurity clearance must be covered by the importer or agent.
- (2) Consignments that do not comply with the requirements of the IHS may be re-shipped or destroyed using an MPI-approved destruction method.

5 Guidance

5.1 Genetically modified (GM) organisms

- (1) Under the Hazardous Substances and New Organisms (HSNO) Act 1996, a GM organism is considered a new organism.
- (2) GM organisms are organisms whose genes or other genetic material have been modified by *in vitro* techniques.
- (3) Organisms that result solely from artificial insemination, superovulation, embryo transfer, or embryo splitting are **not** considered to be GM organisms.
- (4) For more information please visit: <https://www.epa.govt.nz/industry-areas/new-organisms/>.

5.2 Equivalence

- (1) MPI may accept an alternative method, system or process that can be shown to achieve the biosecurity requirements of the IHS (i.e. equivalence).
- (2) MPI's preference is that the exporting country's Competent Authority makes equivalence requests. Equivalence requests can be lodged with animal.imports@mpi.govt.nz.
- (3) An import permit is not required to import equine semen and embryos into New Zealand if the requirements of the IHS are met.
- (4) An import permit may be required where specific equivalence measures are approved by MPI. An import permit serves as evidence of equivalence decisions and will be written as specific notes in the special conditions section of the permit.
- (5) Import permit application forms can be found on the MPI website at [Application form for Semen and Embryos](#).
- (6) Completed applications are lodged with animal.imports@mpi.govt.nz.

5.3 Harmonised system (HS) codes

- (1) The harmonised system is an international product numbering classification developed by the World Customs Organisation (WCO). The New Zealand harmonised system is found here: <https://www.customs.govt.nz/business/tariffs/working-tariff-document/>
- (2) Animal products imported using the IHS will be under one of the following HS Codes:

HS Code	Commodity Description
0511.99.00 08H	Products of animal origin, not elsewhere specified or included; semen, other than bovine, other than sheep semen
0511.99.00 45B	Products of animal origin, not elsewhere specified or included; other, embryos

5.4 Exporting country systems and certification

5.4.1 Approval for exporting systems

- (1) The IHS applies to equine semen and embryos from specified countries. These countries are listed in the IHS. The IHS is based upon an import risk analysis (2009 IRA) written for specified countries which include Australia, Canada, the USA, and the EU. Based on equivalent disease freedom status, Switzerland and Norway are included in the IHS.
- (2) Competent Authorities of other countries must approach MPI for approval of their exporting systems to allow for imports of equine semen and embryos into New Zealand. Approval will also require an import risk analysis to be undertaken for these countries.

5.4.2 Agreed country specific veterinary certificates

- (1) Requests from exporting countries to negotiate veterinary certification for the import of equine semen and embryos into New Zealand will be prioritised according to MPI resources available at the time of application.
- (2) A model veterinary certificate is provided in this guidance document and can be used by the Competent Authority as a reference for country-specific veterinary certificate negotiation.
- (3) All country-specific veterinary certificates agreed between an exporting country's Competent Authority and MPI are included in the table below:

For semen:

Country	Link to certificate	S27 CTO direction #	Date agreed	Date applicable for use
France	Veterinary health certificate for equine semen exported from France to New Zealand	-	30 January 2023	30 January 2023

For embryos:

Country	Link to certificate	S27 CTO direction #	Date agreed	Date applicable for use

- (4) Country-specific veterinary certificates with equivalent measures will be recorded with a number relevant to a Chief Technical Officer (CTO) direction under section 27(1)d(iii) of the Act, to enable border staff to clear the goods and record the number in the MPI database.
- (5) When a newly negotiated country-specific veterinary certificate replaces one which is currently in use, the application of new import conditions will apply according to the dates listed in the table. At that time previous veterinary certificates for that country can no longer be used.
- (6) When a country-specific veterinary certificate is agreed, there will be a four-month transition period to allow consignments of equine semen and embryos to be prepared in accordance with the new conditions. During transition, both the old and the new import conditions are acceptable. After transition, the previous veterinary certificate for that country can no longer be used.

5.5 Diagnostic tests, vaccines and treatment

- (1) MPI lists all approved diagnostic tests and vaccines in the MPI document: [Approved Diagnostic Tests, Vaccines, Treatments and Post-Arrival Testing Laboratories for Animal Import Health Standards, MPI-STD-TVTL](#).
- (2) Where World Organisation of Animal Health (WOAH, founded as OIE) recommended diagnostic tests and vaccines are listed, details can be found in the WOAH *Manual of Diagnostic Tests and Vaccines* found on the WOAH website: <https://www.woah.org/en/what-we-do/standards/codes-and-manuals/terrestrial-manual-online-access/>
- (3) Semen collection may commence upon entry into the semen collection centre and held until all residency and disease testing requirements have been met. Disease testing for semen is valid for semen collected from the date of entry to the semen collection centre until a maximum of 180 days from the date the sample was collected for a single test, or the date of the final test in that series (e.g. CEM) provided:
 - a) The donor remains continuously resident at the semen collection centre;
 - b) The semen collection centre remains under the supervision of the Competent Authority;
 - c) All equids at the semen collection centre remain free from evidence of infectious disease transmissible in equine germplasm;
 - d) While resident at the semen collection centre, donors have only had contact with equids of equivalent tested health status;
 - e) The donors have not been mated by natural service after entering the semen collection centre and during the collection period;

- f) If any of the above provisions cease to apply or if more than 180 days have passed since the sample collection date, the donors and all in contact equids at the semen collection centre must be re-tested in accordance the requirements of the IHS.
- (4) Disease testing for teaser mares is valid until a maximum of 180 days from the date the sample was collected for a single test, or the date of the final test in that series (e.g. CEM) provided:
 - a) The teaser mare remains continuously resident at the semen collection centre;
 - b) The semen collection centre remains under the supervision of the Competent Authority;
 - c) All equids at the semen collection centre remain free from evidence of infectious disease transmissible in equine germplasm;
 - d) While resident at the semen collection centre, teaser mares have only had contact with equids of equivalent tested health status;
 - e) The teaser mares have not been mated by natural service after entering the semen collection centre and during the collection period;
 - f) If any of the above provisions cease to apply or if more than 180 days have passed since the sample collection date, the teaser mares and all in contact equids at the semen collection centre must be re-tested in accordance the requirements of the IHS.

5.6 Inspection and verification

- (1) On arrival, all documentation accompanying the consignment will be verified by an inspector. The inspector may also inspect the consignment, or a sample of the consignment on arrival.
- (2) Inspectors are able to inspect and verify due to their authorised powers under the Act.
- (3) These requirements are independent of the IHS requirements.

6 Specified Requirements for Identified Risk Organisms

6.1 Model veterinary certificate for equine semen

- (1) Below is a model veterinary certificate for trade in equine semen. This model meets the requirements of the IHS.


Part 1: Details of dispatched consignment	1.1. Consignor (Exporter): Name: Address:	1.2. Certificate reference number:													
		1.3. Competent Authority:													
	1.4. Consignee (Importer): Name: Address:														
	1.5. Country of origin: ISO Code**	1.6. Zone or compartment of origin:													
	1.7. Country of destination: ISO Code**	1.8. Zone or compartment of destination:													
	1.9. Place of origin: Name: Address:														
	1.10. Place of shipment:	1.11. Date of departure:													
	1.12. Means of transport: <input type="checkbox"/> Aeroplane <input type="checkbox"/> Ship Identification:	1.13. Expected border post:													
		1.14. CITES permit No(s):													
	1.15. Description of commodity:	1.16. Commodity Code (ISO Code**):													
		1.17. Total number of:													
	1.18. Temperature of commodities for transport: <input type="checkbox"/> Chilled <input type="checkbox"/> Frozen	1.19. Total number of packages:													
1.20. Identification of container/seal number:	1.21. Type of packaging:														
1.22. Identification of commodity: Species (scientific name)***															
<table border="1"> <tr> <td>Donor name</td> <td></td> </tr> <tr> <td>Donor identification</td> <td></td> </tr> <tr> <td>Date of entry into semen collection centre</td> <td>dd/mm/yyyy</td> </tr> <tr> <td>Semen collection period or season</td> <td>dd/mm/yyyy to dd/mm/yyyy</td> </tr> <tr> <td>Date(s) of semen collection</td> <td>dd/mm/yyyy</td> </tr> <tr> <td>Straw/container identification</td> <td></td> </tr> <tr> <td>Number of straws/containers</td> <td></td> </tr> </table>		Donor name		Donor identification		Date of entry into semen collection centre	dd/mm/yyyy	Semen collection period or season	dd/mm/yyyy to dd/mm/yyyy	Date(s) of semen collection	dd/mm/yyyy	Straw/container identification		Number of straws/containers	
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Date(s) of semen collection	dd/mm/yyyy														
Straw/container identification															
Number of straws/containers															
** Optional ***Table does not need to be completed if summary table is used instead															

Part 2: Veterinary Information	Country:	Certificate reference number:
	<p>I, the undersigned Official Veterinarian, certify that the product described above satisfy the following requirements:</p> <p>Eligibility</p> <p>(1) The semen is from horses (<i>Equus caballus</i>) and/or donkeys (<i>Equus asinus</i>).</p> <p>(2) The semen is <i>(delete as applicable)</i>:</p> <p style="margin-left: 40px;">(a) fresh-chilled; or</p> <p style="margin-left: 40px;">(b) frozen</p> <p>(3) The semen is not genetically modified.</p> <p>Diagnostic testing, vaccination, and treatment</p> <p>(4) Diagnostic testing was conducted at a laboratory approved to conduct export testing by the Competent Authority of a country approved to export semen to New Zealand (Australia, Canada, European Union Member States, Norway, Switzerland, or the United States).</p> <p>(5) Laboratory samples were collected, processed, and stored as recommended in the WOA <i>Terrestrial Animal Health Code</i> and/or <i>Manual of Diagnostic Tests and Vaccines for Terrestrial Animals</i>, or as described in <i>MPI-STD-TVTL</i>.</p> <p>(6) Diagnostic test(s) and vaccines were those that have been approved by MPI and documented in <i>MPI-STD-TVTL</i>.</p> <p>(7) Any products and vaccine(s) administered to satisfy import requirements were administered according to the recommendations of the manufacturer in an approved country (Australia, Canada, European Union Member States, Norway, Switzerland, or the United States).</p> <p>(8) Any vaccine(s) administered to satisfy import requirements were either the final dose of a primary vaccination course or the recommended booster to complement the primary course.</p> <p>(9) Vaccine names and date(s) of vaccination are recorded on this veterinary certificate.</p> <p>(10) Original reports, or copies of reports endorsed by the Official Veterinarian, or a tabulated summary endorsed by the Official Veterinarian of all tests and vaccinations are attached to this veterinary certificate.</p> <p>Semen collection centre requirements</p> <p>(11) The semen collection centre meets the relevant articles of the current recommendations of the WOA <i>Terrestrial Animal Health Code</i> chapter <i>General Hygiene in Semen Collection and Processing Centres</i>.</p> <p style="margin-left: 40px;">Name of semen collection centre:</p> <p style="margin-left: 40px;">Address of semen collection centre:</p> <p style="margin-left: 40px;">Date of most recent semen collection centre approval (dd/mm/yyyy):</p> <p style="margin-left: 40px;">Date of semen collection centre approval for semen collected and stored from previous collection periods (dd/mm/yyyy) <i>(delete if not applicable)</i>:</p> <p style="margin-left: 40px;">Approval number of semen collection centre:</p> <p style="margin-left: 40px;">Name of approved veterinarian:</p> <p>Donor requirements</p> <p>(12) While resident at the semen collection centre, donors only had contact with other equids of equivalent health status.</p> <p>(13) On the day of collection, the approved veterinarian ensured by clinical examination including the external reproductive organs, that donors were free from clinical evidence of infectious diseases transmissible in semen.</p>	

	<p>(14) Donors have not been naturally mated since entry into the semen collection centre, unless required for disease testing.</p> <p>(15) The donors were transferred from one approved semen collection centre to another of equivalent health status without isolation or testing and the following occurred (<i>delete entire clause if not applicable</i>):</p> <ul style="list-style-type: none"> (a) Donors were examined, by the approved semen collection centre veterinarian, and showed no clinical sign of disease transmissible in semen on the day of entry into the facility; and (b) Transfer was direct; and (c) Donors were not in direct or indirect contact with animals of a lower health status; and (d) The means of transport used was disinfected before use. <p>(16) The donor has been approved for the <enter years of breeding season> breeding season on <enter date of approval> (<i>applicable to Australia only; delete if not applicable</i>)</p> <p>Teaser mare requirements (<i>delete if not applicable</i>)</p> <p>(17) Teaser mares used for the collection of the semen were of equivalent health status to the donor.</p> <p>(18) Teaser mares were not used as a teaser from the time of sample collection for disease testing until negative disease test results were returned.</p> <p>(19) On the day of collection, the approved veterinarian ensured by clinical examination including the external reproductive organs, that teaser animals were free from clinical evidence of infectious diseases transmissible in semen.</p> <p>Collection, processing, and storage requirements</p> <p>(20) Semen collection, processing, and storage meets the general considerations for hygienic collection and handling of semen specified in the current recommendations of the WOA <i>Terrestrial Animal Health Code</i> chapter <i>Collection and Processing of Bovine, Small Ruminant and Porcine Semen</i>.</p> <p>(21) Antibiotics were added to the semen diluent/extender in generally accepted levels.</p> <p>(22) The cryogenic or cooling agent used in the freezing process, storage, or transport has not been used previously in association with any other product of animal origin.</p> <p>(23) Semen is in straws, or new or disinfected containers which are sealed and tamper-evident, and clearly and permanently marked to identify the donor and the date(s) of collection.</p> <p>(24) Semen was only stored with germplasm that was collected and processed in accordance with the WOA <i>Terrestrial Animal Health Code</i> chapter <i>Collection and Processing of Bovine, Small Ruminant and Porcine Semen</i>, and/or chapter <i>Collection and Processing of In Vivo Derived Embryos from Livestock and Equids</i>, and/or the IETS Manual, and is of equivalent health status.</p> <p>(25) Semen was held in a storage place approved by the Competent Authority until the time of export.</p> <p>(26) The semen in this consignment originates from <approved origin country>, has been stored in <exporting country>, and is accompanied by (<i>delete entire clause if country of origin and export are the same</i>):</p> <ul style="list-style-type: none"> (a) A declaration from the Competent Authority of <exporting country> that links the semen from <origin country> to the semen being exported to New Zealand; and (b) This veterinary certificate certifying that the semen has been stored and transported in <exporting country> in accordance with the requirements of the <i>Import Health Standard for Semen and Embryos from Equids</i>; and (c) Evidence that the semen was collected, processed, and stored in <origin country> in accordance with the requirements of the <i>Import Health Standard for Semen and Embryos from Equids</i> in the form of either: <ul style="list-style-type: none"> (i) A veterinary certificate issued by the Competent Authority of <origin country> certifying that the semen meets the requirements of the <i>Import Health Standard for Semen and Embryos from Equids</i>; or (ii) A letter from the Competent Authority of <origin country> confirming the semen meets the requirements of the <i>Import Health Standard for Semen and Embryos from Equids</i>.
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	<p>Transport requirements</p> <p>(27) The semen for export, all of equivalent health status, was identified and placed under the supervision of the Official Veterinarian in fresh liquid nitrogen in a container(s) which was <i>(delete as applicable)</i>:</p> <ul style="list-style-type: none"> (a) New; or (b) Prior to loading, the shipper was emptied and inspected and any loose straws removed. The shipper, including all surfaces contacting the straws, was disinfected <p>(28) The transport container was sealed by either the approved veterinarian or an Official Veterinarian, using tamper-evident seals.</p> <p>Seal number:</p> <p>(29) The semen was transferred from one transport container to another <i>(delete entire clause if not applicable)</i>.</p> <p>Date of transfer (dd/mm/yyyy):</p> <p>Reason for transfer:</p> <p>Facility moved from:</p> <p>Facility moved to:</p> <p>Veterinarian who performed transfer (name and signature):</p> <p>SPECIFIC REQUIREMENTS FOR IDENTIFIED RISK ORGANISMS:</p> <p>For equine arteritis virus (EVA):</p> <p>(30) Donors were kept for the 28 days prior to semen collection on premises where no equid has shown any clinical sign of EVA during that period and showed no clinical sign of EVA on the day of semen collection; and</p> <ul style="list-style-type: none"> (a) Were subjected to a test for EVA carried out on a single blood sample collected not less than 21 days after entry into the semen collection centre, with negative results; or (b) Were subjected between six and nine months of age to a test for EVA; <ul style="list-style-type: none"> (i) With a negative result and were immediately vaccinated against EVA and regularly revaccinated in accordance with the recommendations of the manufacturer; or (ii) With a positive result, followed at least 14 days later by a second test showing a stable or decreasing antibody titre; and were immediately vaccinated against EVA and regularly revaccinated in accordance with the recommendations of the manufacturer; or (c) Were isolated and not earlier than 7 days of commencing isolation were subjected to a test for EVA on a blood sample with a negative result, immediately vaccinated for EVA, kept for 21 days following vaccination separated from other equids and regularly revaccinated in accordance with the recommendations of the manufacturer; or (d) Were subjected to a test for EVA on a blood sample with a negative result within 14 days prior to semen collection, and had been separated from other equids not of an equivalent EVA status for 14 days prior to blood sampling until the end of semen collection; or (e) Have been subjected to a test for EVA carried out on a blood sample with a positive result and then: either <ul style="list-style-type: none"> (i) Were subsequently test mated to two mares within six months prior to semen collection, which were subjected to two tests for EVA with negative results on blood samples collected at the time of test mating and again 28 days after the test mating; or (ii) Were subjected to a test for EVA with a negative result, carried out on semen collected within six months prior to collection of the semen to be exported; or (iii) Were subjected to a test for EVA with a negative result, carried out on semen collected within six months after the blood sample was collected, then immediately vaccinated, and regularly revaccinated; or (f) For frozen semen, were subjected with negative results either: <ul style="list-style-type: none"> (i) To a test for EVA carried out on a blood sample taken not earlier than 14 days and not later than 12 months after the collection of the semen for export; or
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	<p>(ii) To a test for EVA carried out on an aliquot of the semen collected immediately prior to processing or on an aliquot of semen collected within 14 to 30 days after the first collection of the semen to be exported; and</p> <p><i>(Option (30)(f) applies only to frozen semen. Frozen semen may use options (30)(a)-(f), fresh chilled semen may use options (30)(a)-(e))</i></p> <p>(31) Teaser mares were kept for the 28 days prior to use as a teaser on premises where no equid has shown any clinical sign of EVA during that period and showed no clinical sign of EVA on the day of semen collection (<i>delete entire clause if not applicable</i>); and</p> <p>(a) Were subjected to a test for EVA carried out on a single blood sample, with negative results:</p> <p>(i) Collected not less than 21 days after entry into the semen collection centre, if not a resident at the semen collection centre; or</p> <p>(ii) Collected at the beginning of the breeding season prior to being used for teasing if resident at the semen collection centre.</p> <p>For equine herpesvirus-1 (EHV-1):</p> <p>(32) Donors/teaser mares (<i>delete teaser mare if not applicable</i>):</p> <p>(a) Were kept on premises where no case of EHV-1 has been reported in the 21 days prior to each collection; and</p> <p>(b) Showed no clinical signs of EHV-1 infection on the day of collection and during the 21 days prior to collection.</p> <p>For equine infectious anaemia (EIA):</p> <p>(33) Donors:</p> <p>(a) Were kept on premises where no case of EIA has been reported during the 90 days prior to each collection; and</p> <p>(b) Showed no clinical sign of EIA on the day of each collection; and</p> <p>(c) Were subjected to a test for EIA not less than 21 days after entry into the collection centre, with a negative result.</p> <p>(34) Teaser mares (<i>delete entire clause if not applicable</i>):</p> <p>(a) Were kept on premises where no case of EIA has been reported during the 90 days prior to each collection; and</p> <p>(b) Showed no clinical sign of EIA on the day of each collection; and</p> <p>(c) Were subjected to a test for EIA carried out on a single blood sample with negative results:</p> <p>(i) Collected not less than 21 days after entry into the semen collection centre if not a resident at the semen collection centre; or</p> <p>(ii) Collected at the beginning of each breeding season prior to being used for teasing if resident at the semen collection centre.</p> <p>For <i>Taylorella equigenitalis</i> and <i>Taylorella asinigenitalis</i> (CEM):</p> <p>(35) Donors/teaser mares (<i>delete teaser mares if not applicable</i>) were kept, since birth or for at least the 60 days prior to collection, in a country recognised by MPI as free from CEM, where no case of CEM has been reported in the 2 years prior to export; or</p> <p>(36) Donors/teaser mares (<i>delete teaser mares if not applicable</i>) were kept, since birth or for at least 60 days prior to collection, on premises where no case of CEM has been reported during that time; and</p> <p>(a) Have had no direct or indirect contact with CEM during the 60 days prior to collection; and</p> <p>(b) Showed no clinical signs of CEM on the day of each collection; and</p> <p>(c) Were subjected to a test for CEM not less than 7 days after entry into the semen collection centre, with negative results;</p>
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	<ul style="list-style-type: none"> (i) Stallions were sampled two times at intervals of 4-14 days. Swab sampling sites were the urethra, urethral fossa and its sinus, and the penile sheath. (ii) Teaser mares were sampled two times at intervals of 4-14 days. Swab sampling sites are the clitoral fossa and sinuses (<i>delete if not applicable</i>); and (d) Did not receive antibiotics in the 7 days (systemic treatment) or 21 days (local treatment) before the first sample collection or during the CEM sampling period; and (e) Were protected against any possibility of infection with CEM since the beginning of the tests; or <p>(37) Donors/teaser mares (<i>delete teaser mares if not applicable</i>) have previously shown signs of CEM or have been in direct or indirect contact with CEM during the 60 days prior to collection; and</p> <ul style="list-style-type: none"> (a) Were treated for CEM; and (b) After treatment, were subjected to a test for CEM, with negative results: <ul style="list-style-type: none"> (i) Stallions were sampled three times at intervals of 7-14 days (sampling sites were the urethra, urethral fossa and its sinus, and the penile sheath). Thereafter, the first three mares mated or inseminated by the stallion were tested on clitoral swabs taken 3 times at intervals of at least 7 days, starting 2 days after mating or insemination. (ii) Teaser mares were sampled three times at intervals of 7-14 days (sampling sites are the clitoral fossa and sinuses), and 3 endometrial swabs taken during the next 3 oestrus periods. Maiden mares only required 1 endometrial swab (<i>delete if not applicable</i>); and (c) Were protected against any possibility of infection with CEM since the beginning of the tests. <p>For <i>Trypanosoma equiperdum</i> (dourine):</p> <p>(38) Donors/teaser mares (<i>delete teaser mares if not applicable</i>) were kept since birth, or for the 180 days prior to collection of semen for export, in a country which has been free from dourine for not less than the 180 days prior to export; or</p> <p>(39) Donors were kept for the 180 days prior to collection of semen for export on a premises where no case of dourine was reported during that period; and</p> <ul style="list-style-type: none"> (a) Were subjected to a diagnostic test for dourine with negative results; and (b) The microscopic examination of the semen for dourine was negative. <p>(40) Teaser mares were kept for the 180 days prior to use as a teaser on premises where no case of dourine was reported during that period and were subjected to a diagnostic test for dourine with negative results (<i>delete if not applicable</i>).</p>
<p>Official Veterinarian</p> <p>Name:</p> <p>Address:</p> <p>Email:</p>	<p>Signature:</p> <p>Date (dd/mm/yyyy):</p> <div style="text-align: center;">  <p>Official Veterinarian stamp</p> </div>

This table accompanies the veterinary certificate with reference number: _____

Donor and semen information					
Donor identification	Number of straw/containers	Straw/container identification	Date of entry into semen collection centre	Dates of semen collection period	Date(s) of collection
			dd/mm/yyyy	dd/mm/yyyy to dd/mm/yyyy	dd/mm/yyyy
			dd/mm/yyyy	dd/mm/yyyy to dd/mm/yyyy	dd/mm/yyyy
			dd/mm/yyyy	dd/mm/yyyy to dd/mm/yyyy	dd/mm/yyyy

Test information												
	Equine viral arteritis (EVA)			Equine infectious anaemia (EIA)			Contagious equine metritis (CEM)			Dourine		
Donor/teaser identification	Sample date	Test type	Result	Sample date	Test type	Result	Sample dates	Test type	Result	Sample date	Test type	Result
	dd/mm/yyyy			dd/mm/yyyy			dd/mm/yyyy			dd/mm/yyyy		
	dd/mm/yyyy			dd/mm/yyyy			dd/mm/yyyy			dd/mm/yyyy		

Vaccination information			
Donor identification	Equine viral arteritis (EVA)		
	Primary vaccination date(s)	Booster vaccination date	Vaccine name
	dd/mm/yyyy	dd/mm/yyyy	
	dd/mm/yyyy	dd/mm/yyyy	

Official Veterinarian

Name:

Date:

Signature:



6.2 Model veterinary certificate for equine embryos


- (1) Below is a model veterinary certificate for trade in equine embryos. This model meets the requirements of the IHS.

Part 1: Details of dispatched consignment	1.1. Consignor (Exporter): Name: Address:	1.2. Certificate reference number:											
		1.3. Competent Authority:											
	1.4. Consignee (Importer): Name: Address:												
	1.5. Country of origin: ISO Code**	1.6. Zone or compartment of origin:											
	1.7. Country of destination: ISO Code**	1.8. Zone or compartment of destination:											
	1.9. Place of origin: Name: Address:												
	1.10. Place of shipment:	1.11. Date of departure:											
	1.12. Means of transport: <input type="checkbox"/> Aeroplane <input type="checkbox"/> Ship Identification:	1.13. Expected border post: 1.14. CITES permit No(s):											
	1.15. Description of commodity:	1.16. Commodity Code (ISO Code**): 1.17. Total number of:											
	1.18. Temperature of commodities for transport: <input type="checkbox"/> Chilled <input type="checkbox"/> Frozen	1.19. Total number of packages:											
1.20. Identification of container/seal number:	1.21. Type of packaging:												
1.22. Identification of commodity: (Scientific name)***													
<table border="1"> <tr> <td>Donor name</td> <td></td> </tr> <tr> <td>Donor identification</td> <td></td> </tr> <tr> <td>Date of entry into embryo collection centre or herd</td> <td>dd/mm/yyyy</td> </tr> <tr> <td>Date(s) of embryo collection</td> <td>dd/mm/yyyy</td> </tr> <tr> <td>Straw/container identification</td> <td></td> </tr> <tr> <td>Number of straws/containers</td> <td></td> </tr> </table>		Donor name		Donor identification		Date of entry into embryo collection centre or herd	dd/mm/yyyy	Date(s) of embryo collection	dd/mm/yyyy	Straw/container identification		Number of straws/containers	
Donor name													
Donor identification													
Date of entry into embryo collection centre or herd	dd/mm/yyyy												
Date(s) of embryo collection	dd/mm/yyyy												
Straw/container identification													
Number of straws/containers													
Optional *Table does not need to be completed if summary table is used instead													

Part 2: Veterinary information	Country:	Certificate reference number:
	<p>I, the undersigned Official Veterinarian, certify that the product described above satisfy the following requirements:</p> <p>Eligibility</p> <p>(1) The embryos are from horses (<i>Equus caballus</i>) and/or donkeys (<i>Equus asinus</i>).</p> <p>(2) The embryos are <i>in vivo</i>-derived and frozen</p> <p>(3) The embryos are not genetically modified.</p> <p>Diagnostic testing, vaccination, and treatment</p> <p>(4) Diagnostic testing was conducted at a laboratory approved to conduct export testing by the Competent Authority of a country approved to export embryos to New Zealand (Australia, Canada, European Union Member States, Norway, Switzerland, or the United States).</p> <p>(5) Laboratory samples were collected, processed, and stored as recommended in the WOA <i>Terrestrial Animal Health Code</i> and/or <i>Manual of Diagnostic Tests and Vaccines for Terrestrial Animals</i>, or as described in <i>MPI-STD-TVTL</i>.</p> <p>(6) Diagnostic test(s) and vaccines were those that have been approved by MPI and documented in <i>MPI-STD-TVTL</i>.</p> <p>(7) Any products and vaccine(s) administered to satisfy import requirements were administered according to the recommendations of the manufacturer in an approved country (Australia, Canada, European Union Member States, Norway, Switzerland, or the United States).</p> <p>(8) Any vaccine(s) administered to satisfy import requirements was/were either the final dose of a primary vaccination course or the recommended booster to complement the primary course.</p> <p>(9) Vaccine names and date(s) of vaccination are recorded on this veterinary certificate.</p> <p>(10) Original reports, or copies of reports endorsed by the Official Veterinarian, or a tabulated summary endorsed by the Official Veterinarian of all tests and vaccinations are attached to this veterinary certificate.</p> <p>Embryo collection team requirements</p> <p>(11) The embryo collection team meets the current recommendations of the WOA <i>Terrestrial Animal Health Code</i> chapter <i>Collection and Processing of In Vivo Derived Embryos from Livestock and Equids</i>.</p> <p>Name of embryo collection herd/centre:</p> <p>Address of embryo collection herd/centre:</p> <p>Date(s) of embryo collection team approval for collection period(s) (dd/mm/yyyy):</p> <p>Approval number of embryo collection team:</p> <p>Name of approved veterinarian:</p> <p>Donor requirements</p> <p>(12) Donors were isolated from other equids not of an equivalent tested health status from the time of sample collection for the pre-collection test until completion of embryo collection for export to New Zealand</p> <p>(13) On the day of collection, the approved veterinarian ensured by clinical examination including the external reproductive organs, that donors were free from clinical evidence of infectious diseases transmissible in germplasm.</p> <p>(14) Donors have (<i>delete as applicable</i>):</p> <p>(a) Not been naturally mated since entry into the embryo collection herd/centre, unless required for disease testing.</p>	

	<p>(b) The semen used for the artificial insemination of embryo donors complies with the equine semen requirements of the <i>Import Health Standard for Semen and Embryos from Equids</i>.</p> <p>Collection, processing, and storage requirements</p> <p>(15) Embryo collection, processing, and storage meets the current recommendations of the WOA <i>Terrestrial Animal Health Code</i> chapter <i>Collection and Processing of In Vivo Derived Embryos from Livestock and Equids</i>.</p> <p>(16) Antibiotics were added during collection, processing, washing, and storage media as recommended in the IETS Manual.</p> <p>(17) Embryos had an intact zona pellucida and were free of adherent material after the final wash when examined over its entire surface at not less than 50X magnification. Any micro-manipulation that caused a breach of the zona pellucida, was performed according to the procedures described in the WOA <i>Terrestrial Animal Health Code</i> chapter <i>Collection and Processing of In Vivo Derived Embryos from Livestock and Equids</i> and IETS Manual.</p> <p>(18) The cryogenic or cooling agent used in the freezing process, storage, or transport has not been used previously in association with any other product of animal origin.</p> <p>(19) Embryos are in new or disinfected containers which are sealed and tamper-evident, and clearly and permanently marked to identify the donor and the date(s) of collection. The marking conforms to the international standards of the IETS.</p> <p>(20) Embryos were only stored with germplasm that was collected and processed in accordance with the WOA <i>Terrestrial Animal Health Code</i> chapter <i>Collection and Processing of Bovine, Small Ruminant and Porcine Semen</i>, and/or chapter <i>Collection and Processing of In-Vivo Derived Embryos from Livestock and Equids</i>, and/or the IETS Manual, and is of equivalent health status.</p> <p>(21) Embryos were held in a storage place approved by the Competent Authority until the time of export.</p> <p>(22) The embryos in this consignment originate from <approved origin country>, have been stored in <exporting country>, and are accompanied by (delete entire clause if country of origin and export are the same):</p> <p>(a) A declaration from the Competent Authority of <exporting country> that links the embryos from <origin country> to the embryos being exported to New Zealand; and</p> <p>(b) This veterinary certificate certifying that the embryos have been stored and transported in <exporting country> in accordance with the requirements of the <i>Import Health Standard for Semen and Embryos from Equids</i>; and</p> <p>(c) Evidence that the embryos were collected, processed, and stored in <origin country> in accordance with the requirements of the <i>Import Health Standard for Semen and Embryos from Equids</i> in the form of either:</p> <p>(i) A veterinary certificate issued by the Competent Authority of <origin country> certifying that the embryos meet the requirements of the <i>Import Health Standard for Semen and Embryos from Equids</i>; or</p> <p>(ii) A letter from the Competent Authority of <origin country> confirming the embryos meet the requirements of the <i>Import Health Standard for Semen and Embryos from Equids</i>.</p> <p>Transport requirements</p> <p>(23) The embryos for export, all of equivalent health status, was identified and placed under the supervision of the Official Veterinarian in fresh liquid nitrogen in a container(s) which was (delete as applicable):</p> <p>(a) New; or</p> <p>(b) Prior to loading, the shipper was emptied and inspected and any loose straws removed. The shipper, including all surfaces contacting the straws, was disinfected.</p> <p>(24) The transport container was sealed by either the approved veterinarian or an Official Veterinarian, using tamper-evident seals:</p> <p>Seal number:</p> <p>(25) The embryos were transferred from one transport container to another (delete entire clause if not applicable).</p> <p>Date of transfer (dd/mm/yyyy):</p> <p>Reason for transfer:</p>
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	<p>Facility moved from:</p> <p>Facility moved to:</p> <p>Veterinarian who performed transfer (name and signature):</p> <p>SPECIFIC REQUIREMENTS FOR IDENTIFIED RISK ORGANISMS:</p> <p>For equine arteritis virus (EVA):</p> <p>(26) Donors showed no clinical sign of EVA on the day of embryo collection; and</p> <p>(a) Were kept for the 28 days prior to embryo collection on premises where no equid has shown any clinical sign of EVA during that period; and</p> <p>(i) Were subjected to a test for EVA carried out on a single blood sample collected either once within 21 days prior to collection with negative results, or on two occasions at least 14 days apart within 28 days prior to collection, which demonstrated stable or declining antibody titres; or</p> <p>(ii) Were regularly vaccinated in accordance with the recommendations of the manufacturer; or</p> <p>(b) Donors were isolated for the 28 days prior to collection and have not shown any clinical signs of EVA during that period.</p> <p>For equine herpesvirus-1 (EHV-1):</p> <p>(27) Donors:</p> <p>(a) Were kept on premises where no case of EHV-1 has been reported in the 21 days prior to each collection; and</p> <p>(b) Showed no clinical signs of EHV-1 infection on the day of collection and during the 21 days prior to collection.</p> <p>For equine infectious anaemia (EIA):</p> <p>(28) Donors:</p> <p>(a) Were kept on premises where no case of EIA has been reported during the 90 days prior to and the 60 days after each collection; and</p> <p>(b) Showed no clinical sign of EIA on the day of each collection; and</p> <p>(c) Were subjected to a test for EIA in the 30-60 days after collection, with a negative result.</p> <p>For <i>Taylorella equigenitalis</i> and <i>Taylorella asinigenitalis</i> (CEM):</p> <p>(29) Donors were kept, since birth or for at least the 60 days prior to collection, in a country recognised by MPI as free from CEM, where no case of CEM has been reported in the 2 years prior to export; or</p> <p>(30) Donors were kept, since birth or for at least 60 days prior to collection, on premises where no case of CEM has been reported during that time; and</p> <p>(a) Have had no direct or indirect contact with CEM during the two months prior to collection; and</p> <p>(b) Showed no clinical signs of CEM on the day of each collection; and</p> <p>(c) Were subjected to a test for CEM within 30 days prior to collection with negative results. Donors were sampled two times at intervals of 4-14 days (swab sampling sites are the clitoral fossa and sinuses); and</p> <p>(d) Did not receive antibiotics in the 7 days (systemic treatment) or 21 days (local treatment) before the first sample collection or during the CEM sampling period; or</p> <p>(31) Donors have previously shown signs of CEM or have been in direct or indirect contact with CEM during the 60 days prior to collection; and</p> <p>(a) Were treated for CEM; and</p>
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	<p>(b) After treatment, were subjected to a test for CEM, with negative results. Donors were sampled three times at intervals of 7-14 days (sampling sites are the clitoral fossa and sinuses), and 3 endometrial swabs taken during the next 3 oestrus periods. Maiden mares only require 1 endometrial swab; and</p> <p>(c) Were protected against any possibility of infection with CEM since the beginning of the tests.</p> <p>For <i>Trypanosoma equiperdum</i> (dourine):</p> <p>(32) Donors were kept since birth, or for the 180 days prior to collection of embryos for export, in a country which has been free from dourine for not less than the 180 days prior to export; or</p> <p>(33) Donors were kept for the 180 days prior to collection of semen for export on a premises where no case of dourine was reported during that period and were subjected to a diagnostic test for dourine with negative results.</p>	
	<p>Official Veterinarian</p> <p>Name:</p> <p>Address:</p> <p>Email:</p>	<p>Signature:</p> <p>Date (dd/mm/yyyy):</p> <div data-bbox="1209 824 1391 1003" style="text-align: center;">  <p>Official Veterinarian stamp</p> </div>

This table accompanies the veterinary certificate with reference number: _____

Donor information							
Female donor identification	Number of containers	Number of embryos/container	Container identification	Date of entry into embryo collection herd/centre	Date(s) of collection	Male donor identification	Date of semen collection/natural mating
				dd/mm/yyyy	dd/mm/yyyy		dd/mm/yyyy
				dd/mm/yyyy	dd/mm/yyyy		dd/mm/yyyy
				dd/mm/yyyy	dd/mm/yyyy		dd/mm/yyyy

Test information												
	Equine viral arteritis (EVA)			Equine infectious anaemia (EIA)			Contagious equine metritis (CEM)			Dourine		
Donor identification	Sample date	Test type	Result	Sample date	Test type	Result	Sample dates	Test type	Result	Sample date	Test type	Result
	dd/mm/yyyy			dd/mm/yyyy			dd/mm/yyyy			dd/mm/yyyy		
	dd/mm/yyyy			dd/mm/yyyy			dd/mm/yyyy			dd/mm/yyyy		

Vaccination information			
	Equine viral arteritis (EVA)		
Donor identification	Primary vaccination date(s)	Booster vaccination date	Vaccine name
	dd/mm/yyyy	dd/mm/yyyy	
	dd/mm/yyyy	dd/mm/yyyy	

Official Veterinarian

Name:

Date:

Signature:



Appendix 1 – Document History

Date First Issued	Title	Shortcode
3 December 2015	Guidance Document: Semen and Embryos from Horses (<i>Equidae</i>)	HORSSEMB.SPE
Date of Issued Amendments	Title	Shortcode
7 July 2017	Guidance Document: Semen and Embryos from Horses (<i>Equidae</i>)	HORSSEMB.SPE
9 May 2018	Guidance Document: Semen and Embryos from Horses (<i>Equidae</i>)	HORSSEMB.SPE
5 September 2019	Guidance Document: Semen and Embryos from Equids	EQUIGERM.SPE
15 March 2023	Guidance Document: Semen and Embryos from Equids	EQUIGERM.SPE