Ministry for Primary Industries
Manatū Ahu Matua

Animal Products Notice

Day-Old Chicks and Hatching Eggs – Additional Disease Requirements for Specified Markets

19DXP

Effective from 8 July 2022

Issued under the Animal Products Act 1999

New Zealand Government

TITLE

Animal Products Notice: Day-Old Chicks and Hatching Eggs – Additional Disease Requirements for Specified Markets

COMMENCEMENT

This Animal Products Notice comes into force on date of signing.

REVOCATION

This Animal Products Notice revokes and replaces:

- Day-Old Chicks and Hatching Eggs – Additional Disease Requirements for Specified Markets, dated 24 February 2022

ISSUING AUTHORITY

This Animal Products Notice is issued under sections 167(1) and 60(1) of the Animal Products Act 1999.

Dated at Wellington, 8 July 2022

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Ministry for Primary Industries
(acting under delegated authority of the Director-General)

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Introduction

This introduction is not part of the Animal Products Notice, but is intended to indicate its general effect.

Purpose

The purpose of this document is to set out additional zoosanitary requirements necessary to export compliant day-old chicks and hatching eggs from New Zealand to the countries listed in Schedule 1 of this Notice.

Background

The Animal Products Act 1999 provides the controls and mechanisms needed to give and to safeguard official assurances or zoosanitary certificates to facilitate the entry of animal material including live animals, hatching eggs, semen and embryos, and products into overseas markets.

Notices issued as Overseas Market Access Requirements (OMARs) under the Animal Products Act specify the requirements that are necessary or desirable for the purpose of facilitating access to overseas markets or are in accordance with the requirements of the relevant authority of the importing country. This OMAR safeguards the assurances provided by New Zealand for specific diseases, and should be read in conjunction with the specific OMARS.

Up until recently, *Salmonella* Enteritidis had not been detected in New Zealand commercial poultry flocks. As a result of the recent finding of positive results from several poultry farms, the current controls, verification, and testing levels are required to be strengthened to meet the *Salmonella* Enteritidis expectations of the specified markets.

This OMAR notification has been updated to modify testing requirements and to better align with the Emergency Control Scheme.

Who should read this Animal Products Notice?

Exporters of day-old chicks and hatching eggs to those countries specified in Schedule 1 of this Notice.

Operators of Export Approved Premises collecting day-old chicks and hatching eggs for export to those countries specified in Schedule 1 of this Notice.

Recognised agencies and recognised persons who undertake verification activities and laboratory testing for the export of day-old chicks and hatching eggs to which this Notice applies.

Why is this important?

This Notice is important because it sets out the requirements that need to be met so that the Director-General of the New Zealand Ministry for Primary Industries (MPI) can certify that day-old chicks and hatching eggs meet the requirements for export to the countries listed in Schedule 1 of this Notice, which New Zealand has determined will apply. It should be noted that although the day-old chicks and hatching eggs may comply with these requirements and be given an official assurance (by way of a certificate), the importing country ultimately retains control over what product it clears for entry.

Document History

Version Date	Section Changed	Change(s) Description
9 July 2021	All sections	Additional disease requirements for multiple markets.
3 September 2021	1.3 (4) 1.4 & 1.6 1.4 (3) a) & b) Schedule 2 (3) d) Schedule 4, Dust Sampling	Clarification around loss of export eligibility. Removal of superfluous date references. Clarification in relation to final moisture content and inactivation treatment and the addition of equivalence option. Clarification of dust swab compositing. Change reference to fan belts, use fan grills instead.
24 February 2022	1.4 (4) c) 1.6 (4) Schedule 2 (3) a) Schedule 4, boot sampling	Change to wording around frequency of testing of source flocks in lay. Addition of source farms for verification and change to the frequency of verification. Change to the minimum sampling requirements and addition of a guidance box. Change to the number of boot swabs.
8 July 2022	1.4 (4) (c) Schedule 2 (3) a)	Change to wording around frequency of testing of source flocks in lay. Change to the minimum sampling requirements.

Other information

Guidance

The information in italics contained within a border throughout this OMAR is for guidance and is not part of the statutory requirements.

Export non-conformances

Exporters should note that, under section 51 of the Animal Products Act 1999, where they have exported animal material or products, including live animals, hatching eggs, semen and embryos, that are refused entry by the foreign government they have a statutory duty to notify the Director-General of MPI not later than 24 hours after they have first knowledge of the event.

Liability

Section 61A of the Animal Products Act 1999 states that:

The Crown is not liable, and nor is the Director-General or any employee of the Ministry liable, for any loss arising through the refusal or failure of the relevant authority of an overseas market to admit export animal material or animal product to that market.

Related documents

OMAR documents can be downloaded from https;//www.mpi.govt.nz/export/omars/omars-live-animals-semen-embryos-organics/

When you click on the + symbol on the right-hand side of any OMAR document, you can view the related information and documents (guidance document and export certificate template).

This should be read in conjunction with market specific OMARs which can be found at the link above.

Part 1: Requirements

1.1 Application

(1) This Notice applies to:

a) Day-old chicks and hatching eggs when exported to the markets listed in Schedule 1.
b) Recognised Agencies and Recognised Persons who undertake verification activities and laboratory testing for the export of day-old chicks and hatching eggs to which this Notice applies.

1.2 Definitions

(1) Any term or expression that is defined in the Animal Products Act 1999 but is not defined in this document has the same meaning as in the Act.

(2) Terms not defined in this Notice have the same meaning as those set out in the relevant country OMARs.

(3) For the purposes of this Notice:

Act means the Animal Products Act 1999

DOC/HE means day-old chicks/hatching eggs

EAP means egg collection and storage facilities, and hatcheries that are Export Approved Premises and supply day-old chicks and hatching eggs for export with an official assurance

Environmental testing means environmental sampling and testing of EAPs and source farms as per Schedule 2

ESR means Environmental Science Research laboratory

Export certificate is the form of an official assurance determined by the Director-General pursuant to section 62 of the Act

Hatchery means a demarcated premise which includes a building containing one or more incubators to incubate and hatch eggs to produce day-old chicks

Non-negative result means a result that is not negative and includes initial test results that subsequently may be confirmed as negative or positive

Overseas Market Access Requirement (OMAR) is an export requirement specified by the Director-General by notice issued under section 167(1) of the Animal Products Act 1999 for the purposes of Section 60 of the Act that specifies requirements for animal material or animal product intended for export

PCR means a polymerase chain reaction test that is performed on a sample that has undergone primary enrichment

Shed means each building capable of containing a discreet population of live chickens (known as a flock) that do not physically mix or share interior airspace with another population (flock) of chickens

Source farm means a demarcated premise in which breeding poultry comprising one or more source flocks are housed in one or more houses/sheds. A source farm will have a single entrance for personnel, equipment, and feed, and may include egg fumigation and storage facilities

Source flock means parent flocks of day-old chicks and/or hatching eggs for export

(4) A term used in this Notice that is defined in the Act or the following Notices (or their successors) has the meaning given to it in the Act or that Notice:

a) *Animal Products Notice: Official Assurances Specifications for Animal Material and Animal Products.*
b) *Animal Products Notice: Specifications for Laboratories.*
c) *Animal Products Notice: Export Approved Premises*.

1.3 *Salmonella* Enteritidis freedom status requirements

(1) EAPs and source farms must be demonstrated free of *Salmonella* Enteritidis in accordance with the requirements of section 1.3 and section 1.4.

(2) The EAP and/or source farm's *Salmonella* Enteritidis freedom status is supported by a documented programme sufficient to demonstrate freedom.

(3) EAPs and source farms must be epidemiologically isolated from any other poultry farm or EAP of unknown or confirmed positive *Salmonella* Enteritidis status, using biosecurity controls for pathogen management set out in Schedule 3.

(4) Any EAP or source farm that returns a confirmed positive result for *Salmonella* Enteritidis loses export eligibility of DOC/HEs for markets in Schedule 1, until the EAP or source farm is confirmed free from *Salmonella* Enteritidis.

(5) For the avoidance of doubt, where any country specific OMAR or relevant Export Certificate stipulates a specified duration of freedom prior to export, then this time period shall be the minimum period before the EAP and/or source farm regains eligibility for that country.

1.4 *Salmonella* testing requirements

(1) An EAP and/or source farm must have testing carried out in accordance with Section 1.4 and Schedule 2.

(2) All tests must be carried out at a MPI Recognised Laboratory and conducted in accordance with the *Animal Products Notice: Specifications for Laboratories*.

(3) Poultry feed used at a source farm must be negative for *Salmonella* in accordance with one of the following:

a) Extruded pellet feed with a final moisture content of 8-13% must undergo a *Salmonella* inactivation treatment of at least 80°C for 60 seconds, or equivalent;
b) Mash feed, extruded or press-pelleted feed that does not meet the parameters of section 1.4 (3) a) above, and any other type of feed, must be sampled and tested in accordance with Schedule 2.

(4) Testing of EAPs and associated source farms must include the following as a minimum:

a) **Hatcheries** must conduct post-hatch testing in accordance with Schedule 2:

i) at least once a week;
ii) or if it has been greater than one week since the last hatch or there is less than one hatch per week, immediately after the next hatch;

b) For **source flocks in rearing,** the rearing shed must be tested in the two weeks:

i) before the flock is moved to the laying shed, or
ii) before the estimated point of lay for birds that remain in the same shed for laying;

c) For **source flocks in lay,** the laying shed must be tested prior to any hatching eggs being prepared for export or set for incubation in an EAP, then within five (5) weeks and every five (5) weeks thereafter.
d) Every shed on a source farm containing a source flock must be tested for the farm to be deemed negative for *Salmonella* Enteritidis.

(5) For the avoidance of doubt, where any country specific OMAR or relevant Export Certificate stipulates a more intensive frequency of testing or a specific type of test, then the requirements for that OMAR or Export Certificate apply for export eligibility for that country.

1.5 Consequences of a non-negative or positive *Salmonella* Enteritidis result

(1) Where any sample returns a non-negative result on PCR for *Salmonella* Enteritidis or where *Salmonella* Enteritidis cannot be excluded:

a) the EAP must immediately notify this to the relevant Recognised Agency; and
b) the Recognised Agency must immediately notify MPI Animal Exports (animalexports@mpi.govt.nz).

(2) The Recognised Laboratory must forward the sample to ESR for further investigation.

(3) Where *Salmonella* Enteritidis has been detected in a source flock, environmental testing of the empty shed must be carried out after depopulation, cleaning and disinfection of the shed.

1.6 Verification

(1) The EAP must ensure verification in relation to the requirements of this Notice is completed as part of normal performance-based verification. Verification must be carried out by a Recognised Person.

(2) Following a confirmed positive detection of *Salmonella* Enteritidis, EAPs and source farms must undertake the requirements of section 1.3 (3), (4) and (5), and be verified by the Recognised Person, prior to submitting an application to regain export eligibility to MPI. Application to be sent to MPI Animal Exports (animal.exports@mpi.govt.nz).

(3) Following verification according to 1.6 (1), verification of the EAP, including source farms, must be completed on site by the Recognised Person at least twice a year.

Schedule 1 – Specified markets

Country	**Diseases requiring additional assurances**
Fiji	*Salmonella* Enteritidis
French Polynesia	*Salmonella* Enteritidis
India	*Salmonella* Enteritidis
Indonesia	*Salmonella* Enteritidis
Japan	*Salmonella* Enteritidis
Malaysia	*Salmonella* Enteritidis
Mauritius	*Salmonella* Enteritidis
Myanmar	*Salmonella* Enteritidis
Nepal	*Salmonella* Enteritidis
New Caledonia	*Salmonella* Enteritidis
Pakistan	*Salmonella* Enteritidis
Sri Lanka	*Salmonella* Enteritidis
Thailand	*Salmonella* Enteritidis
United States of America	*Salmonella* Enteritidis
Vietnam	*Salmonella* Enteritidis

Schedule 2 – Specific testing requirements

(1) Minimum sampling requirements for poultry feed

a) Finished poultry feed that has not undergone a pathogen inactivation treatment must be sampled and tested with negative *Salmonella* results. This may be demonstrated in the following manner, or via an alternative method approved by the Recognised Agency as meeting alternative MPI criteria:

i) For feed made up on site, at least five random 25g grab samples, which can be composited, must be tested from each finished batch;
ii) For feed brought in, suppliers must demonstrate evidence of a negative *Salmonella* test for each batch, or it must be tested as per clause i) above.

(2) Minimum sampling requirements for the hatchery

a) Post-hatch testing at the hatchery of the following as appropriate (see guidance on sample methods in Schedule 4):

i) Chick paper / meconium swabs;
ii) Chick fluff;
iii) Hatcher tray swabs;
iv) Chick belt swabs;
v) Additional sampling may also include, where possible, testing of dead rodents, rodent droppings and swabs of rodent traps.

A non-negative result for any additional sampling from within the hatchery building or source farm shed should be managed according to section 1.5. Additional sampling outside of the hatchery building or source flock shed is encouraged but results will not affect the eligibility of the DOC/HE.

(3) Minimum sampling requirements for sheds

a) Environmental sampling on each shed must be conducted as follows:

Shed Length	Sample Type	
	Boot Swabs	**Dust Swab or Dust Sample**
Up to 25m	One boot swab pair	One dust swab or one dust sample
25m – 75m	Two boot swab pairs	Two dust swabs or two dust samples
>75m	Four boot swab pairs	Four dust swabs or four dust samples

b) Samples must be representative of the entire shed (see Schedule 4 for sampling guidance).
c) Environmental sampling may also include, where possible, testing of dead rodents, rodent droppings and swabs of rodent traps.

Boot swabs: *For routine sampling, boot swabs from a single shed may be combined into a single sample bag by the operator, up to a maximum of 8 boot swabs (4 pairs) per shed. Operators may choose to keep boot swab pairs separate to give more granularity to the results.*

Dust swabs: *For routine sampling, dust swabs from a single shed may be combined into a single sample bag by the operator, up to a maximum of 8 swabs per shed. Sufficient steady pressure should be applied when sampling with dust swabs to ensure that the sample picks up the dust present.*

Dust samples: *A dust sample consists of 25 g of dust, which may be collected from multiple locations in the shed where significant dust levels are present. For areas where there is little dust present, a dust swab may be substituted.*

If the sampling requirements are unclear, refer to your verifier for guidance.

Dust sampling by ***dust collection*** *is the most effective method of environmental sampling. Dust collections can't be composited but a single dust sample (25g) can be collected from several areas within the shed. Dust swabs should be a large gauze swab, or pre-moistened drag swab, or foam swab stick lollipop type, or equivalent, dependant on availability and sampling site suitability.*

(4) Testing methods

a) Acceptable testing methods:

i) Conventional culture methods with pre-enrichment, selective enrichment and selective plating. Presumptive isolates of *Salmonella* must be treated as non-negative;

ii) PCR screen after primary enrichment for *Salmonella* and *Salmonella* Enteritidis. A sample negative by PCR screen for *Salmonella* may be reported as negative;

iii) All isolates of *Salmonella* Enteritidis, or any *Salmonella* isolates where *Salmonella* Enteritidis cannot be excluded, must be forwarded to ESR for further investigation and serotyping as necessary.

Schedule 3 – Pathogen management

(1) Requirements for a EAP and/or source farm to be epidemiologically isolated include a documented programme that addresses the following biosecurity controls as a minimum:

a) How the *Salmonella* Enteritidis freedom status of any DOC/HEs arriving at the EAP or source farm is confirmed.

b) Appropriate separation, or effective sanitation procedures, controls and testing that address possible introduction of pathogens associated with:

i) human traffic;
ii) vehicular traffic;
iii) equipment use;
iv) stock movement;
v) litter and dead birds;
vi) inputs (e.g. feed, water);
vii) staff;
viii) rodents, insects and wild birds.

Schedule 4 – Sampling methods

Below is recommended as best practice for environmental sampling methods. Alternative sampling can be done but it is recommended this is confirmed as appropriate by the Recognised Agency or Recognised Person responsible for verification of the source farm. Methods used should be included in the farm's documented programme as required by clause 1.3 (2).

(1) Prior to entering the property, sampling personnel should ensure they have all the necessary equipment. Always have clean, disinfected hands before commencing and after any sampling procedure.

(2) The sample bag should be labelled appropriately:

- *a) Date/time collected;*
- *b) Farm name and EAP ID;*
- *c) Shed name/number;*
- *d) Breed, age, and ID of flock;*

(3) The sample should be stored and sent to the lab chilled (not frozen).

Boot Swab Sampling

(1) Materials needed (per shed)

- *a) Boot swab kit – pre-moistened cotton-poly blend fabric sock style boot;*
- *b) Plastic boot cover (to wear under the pre-moistened boot swab);*
- *c) Disposable latex gloves;*
- *d) Original twist tie bag the boot swabs came in;*
- *e) Gauze swabs or pre-moistened drag swabs for dust samples (if dust sparse – otherwise hand sweep collects are preferable) or foam swab lollipop type;*
- *f) Permanent marker and pen;*
- *g) Laboratory submission form;*
- *h) Sample submitters form;*
- *i) Long stick holder fastening swab or foam swab lollipop type (sampling conveyor belts and high surfaces) if required;*
- *j) Cable ties to attach boot swabs to sticks – cable tie cutter (or may be able to simply slide off).*

(2) All boot swabs must be handled with clean disposable gloves to prevent cross-contamination. Wearing a disposable glove, slip on the disposable plastic boot cover to prevent cross-contamination. It is very important that the boot swab does not come into contact with any sanitation fluid that may have been used on boots.

(3) Open twist tie bag and remove the pre-moistened boot swab and place it securely over your boot and plastic boot cover as per Figure 1.

Figure 1: Boot swab set-up

(4) Boot swabs must be worn whilst the dust sampling tasks are being carried out. Smaller sheds are sampled with one pair of boot swabs, larger sheds are sampled with four pairs of boot swabs (1 pair for each quarter). In addition to the area covered during the dust sampling ensure a path is taken through the shed area similar to the path in Figure 2. (below) or as appropriate to ensure sufficient coverage of the entire floor area.

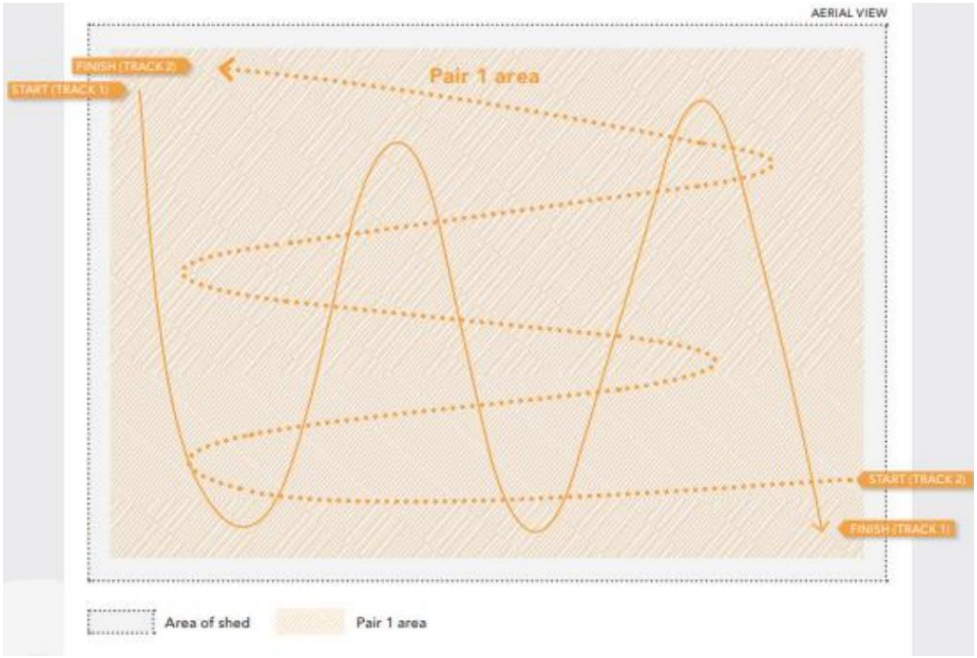

Figure 2: Swabbing pattern for boot swabs

(5) Remove the boot swab from boot, return it to its original bag and seal the bag using the twist tie. A pair of boot swabs can be put in a single sample bag which will constitute one boot swab sample.

Dust Sampling

(1) Salmonella can survive in dust for long periods. Try to choose a good sampling site where dust is observed, including ledges, fan grills, egg belts/slats, feeders, water lines, walls, nest boxes and vents.

(2) Materials needed (per shed):

- *a) Disposable latex gloves;*
- *b) Original twist tie bag;*
- *c) Permanent marker;*
- *d) Laboratory submission form.*

(3) Additional materials for the alternative dust swab method:

- *a) Pre-moistened drag swab(s);*
- *b) Original pottle or twist tie bag.*

(4) Preferred method for dust sampling:

- *a) Wear the disposable glove on your hand to collect the sample;*
- *b) Open the original pottle or twist tie bag;*
- *c) Using your gloved hand, scrape dust from surfaces into the sample bag (not from lighting or feed troughs);*
- *d) Aim to collect two tablespoons in volume (approximately 25g) from at least 1 square meter of surfaces.*

(5) Alternative dust swab for sheds with minimal dust levels:

a) Use a fresh set of disposable gloves per sample;
b) Remove the pre-moistened swab from its pottle or twist tie bag;
c) Coat both sides of the swab with dust collected from multiple places in the shed (not from lighting or feed troughs);
d) At least 1 square meter of surface should be covered per swab;
e) Place swab back in the pottle or twist tie bag.

(6) Fold the top of the bag 2-3 times and secure closed with metal tabs to seal or replace the pottle lid.

Hatchery sampling

(1) For the sampling of meconium from chick papers, the chicks should have been in the boxes for at least an hour prior to sampling. Chick papers can be obtained from boxes containing cull chicks prior to disposal. Use a moist tampon drag swab or boot swab to thoroughly swab the surface of ten chick box papers (approximately 1000 chicks).

(2) A boot swab can be used to gather the chick fluff for placing in a sealed bag and delivery to the laboratory. This could reduce the potential of cross contamination in the laboratory.

(3) The hatcher and chick belt can be sampled by using the alternative dust swab method above in "Dust sampling" clause (5).