



# Guidance Document

## **Semen and Embryos from Sheep and Goats**

OVCAGERM.GEN

Draft

## Title

Guidance Document: Semen and Embryos from Sheep and Goats

## About this document

This guidance document contains information about acceptable ways of ensuring compliance with the requirements in the *Import Health Standard (IHS): Semen and Embryos from Sheep (Ovis aries) and Goats (Capra hircus)*.

Any guidance on how to comply with the applicable requirements may not be the only way to achieve compliance. Stakeholders are encouraged to discuss departures from the approaches outlined in this guidance document with the Ministry for Primary Industries (MPI) to avoid expending resources on the development of alternative approaches which may later be considered unsuitable.

The term “must” is not typically used in guidance. In this particular document if the term “must” is used, it is used in the context of quoting or paraphrasing the requirements set out in the *IHS: Semen and Embryos from Sheep (Ovis aries) and Goats (Capra hircus)*.

## Related Requirements

## Change history

Refer to Appendix 1.

## Contact Details

For further information and questions about this guidance document, please contact:

Ministry for Primary Industries  
Animal Imports  
PO Box 2526  
Wellington 6140

Email: [animalimports@mpi.govt.nz](mailto:animalimports@mpi.govt.nz)

## Disclaimer

This guidance does not constitute, and should not be regarded as, legal advice. While every effort has been made to ensure the information in this guidance is accurate, the Ministry for Primary Industries does not accept any responsibility or liability whatsoever for any error of fact, omission, interpretation or opinion that may be present, however it may have occurred.

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# 1 Purpose

- (1) This guidance document has been issued to accompany the *IHS: Semen and Embryos from Sheep (Ovis aries) and Goats (Capra hircus)*.
- (2) This guidance document should be read in conjunction with that standard.
- (3) This document includes:
  - a) A table listing countries with MPI approved export systems to import semen and embryos from sheep (*Ovis aries*) and goats (*Capra hircus*) into New Zealand
  - b) Model semen and embryo Veterinary Certificates
  - c) Negotiated country specific Veterinary Certificates

# 2 Background

- (1) The *IHS: Semen and Embryos from Sheep (Ovis aries) and Goats (Capra hircus)* which this Guidance Document accompanies, contains generic import requirements. These are the rules to manage the biosecurity risk of importing semen and embryos from sheep and goats from all countries that can meet the requirements of the IHS and in doing so meet New Zealand's appropriate level of protection. The generic IHS serves as the basis for country-to-country (bilateral) negotiations countries. This Guidance Document contains a model Veterinary Certificate and the bilaterally-agreed Veterinary Certificate for trade in semen and embryos from sheep (*Ovis aries*) and goats (*Capra hircus*). The country-specific Veterinary Certificates represent what will be certified prior to exporting consignments of semen and embryos from sheep and goats from the countries specified.
- (2) General information about importing germplasm can be found here:  
<http://www.biosecurity.govt.nz/imports/animals/standards/general-info-germplasm.htm>

# 3 Definitions

- (1) Refer to Appendix 1 of the *IHS: Semen and Embryos from Sheep (Ovis aries) and Goats (Capra hircus)*.

# 4 Importer Responsibilities

- (1) The costs to MPI in performing functions relating to the importation of semen and embryos from sheep and goats will be recovered in accordance with the Biosecurity Act 1993 (the Act) and any regulations made under that Act. All costs involved with documentation, transport, storage and obtaining a biosecurity clearance will be covered by the importer or agent.

# 5 Guidance

## 5.1 Equivalence and permits

- (1) MPI may accept an alternative method, system or process that can be shown to achieve the biosecurity requirements of the IHS (i.e. equivalence).
- (2) MPI's preference is that the exporting country's Competent Authority makes equivalence requests. Equivalence requests can be lodged with [animalimports@mpi.govt.nz](mailto:animalimports@mpi.govt.nz)
- (3) Permit to import application forms can be found on the MPI website at:  
<http://www.biosecurity.govt.nz/regs/imports/animals/forms>.

- (4) Where specific equivalence measures are approved by MPI as per the equivalence clause in the IHS, the special condition section of the permit to import serves as evidence of those equivalence decisions.
- (5) Completed applications are lodged with animal imports [animalimports@mpi.govt.nz](mailto:animalimports@mpi.govt.nz).

## 5.2 Incorporation of material by reference

- (1) Incorporation by reference means that standards, guidelines or lists are incorporated into the IHS and they form part of the requirements. This is done because technical documents are too large or impractical to include in the IHS.
- (2) Where the IHS states that section 142O(1) of the Act does not apply, this means that importers need to refer to the most recent version of any standards, guidelines or lists that are incorporated by reference in the IHS.

## 5.3 Inspection and verification

- (1) On arrival, all documentation accompanying the consignment will be verified by an inspector. The inspector may also inspect the consignment, or a sample of the consignment on arrival.
- (2) Inspectors are able to inspect and verify due to their authorised powers under the Act.
- (3) These requirements are independent of the IHS requirements.

## 5.4 Exporting country systems and certification

- (1) Requests from exporting countries to negotiate veterinary certification to export semen and embryos from sheep (*Ovis aries*) and goats (*Capra hircus*) to New Zealand will be prioritised according to MPI resources available at the time of application.
- (2) MPI recommends Competent Authorities refer to the OIE Code section titled "Quality of Veterinary Services," to prepare evidence for MPI regarding capabilities and preferences of the exporting country's Competent Authority.
- (3) A model veterinary certificate is provided in this guidance document and can be used by the Competent Authority as a reference for country-specific veterinary certificate negotiation.
- (4) The table below lists those exporting countries that satisfy the exporting country systems and certification requirements of the IHS.

Countries with Approved Exporting Systems	Date Agreed
Australia	March 2004

## 5.5 Diagnostic test(s) and vaccines for international trade

- (1) MPI lists all approved tests and vaccines in the MPI document: Approved diagnostic tests, vaccines, treatments and post-arrival testing laboratories for animal import health standards (MPI-STD-TVTL).
- (2) The OIE Manual of Diagnostic Tests and Vaccines describes can be found on the OIE website: <http://www.oie.int/en/international-standard-setting/terrestrial-manual/access-online/>.

## 5.6 Agreed country specific Veterinary Certificates

- (1) After issue of the IHS, the IHS measures may be used by countries which already have agreed Veterinary Certificates. Using the measures before a new country specific Veterinary Certificate is agreed can create challenges at the time of biosecurity clearance. MPI should be notified prior to their use in order to provide clarification to border staff.
- (2) All country specific Veterinary Certificates agreed between an exporting country's Competent Authority and MPI are included in the tables below:

For semen:

Country	Link to certificate	S27 CTO direction #	Date agreed (transition begins)	End of transition

For embryos:

Country	Link to certificate	S27 CTO direction #	Date agreed (transition begins)	End of transition

- (3) Country-specific veterinary certificates with equivalent measures will be recorded with a Chief Technical Officer (CTO) direction number to enable border staff to clear the goods and record the number in the MPI database.
- (4) When a country-specific veterinary certificate is agreed, there will be a four month transition period to allow donors to be prepared in accordance with the new conditions. During transition, both the old and the new import conditions are acceptable. After transition, the previous veterinary certificate for that country can no longer be used.

## 5.7 Model Veterinary Certificate for International Trade in Semen from Sheep (*Ovis aries*) and Goats (*Capra hircus*)

- (1) Below is the model veterinary certificate for trade in semen from sheep (*Ovis aries*) and goats (*Capra hircus*). The model meets the requirements of the IHS.
- (2) The model veterinary certificate format is based on the OIE *Code* Chapter for model veterinary certificates for international trade in germplasm.

COUNTRIES:			
Part I: Details of dispatched consignment	I.1. Consignor (Exporter): Name: Address:		I.2. Certificate reference number: I.3. Competent Authority:
	I.4. Consignee (Importer): Name: Address:		
	I.5. Country of origin ISO Code*		I.6. Zone or compartment of origin**:
	I.7. Country of destination: ISO Code*		I.8. Zone or compartment of destination**:
	I.9. Place of origin: Name: Address:		
	I.10. Place of shipment:		I.11. Date of departure:
	I.12. Means of transport: Aeroplane <input type="checkbox"/> Ship <input type="checkbox"/> Identification:		I.13. Expected border post: I.14. CITES permit No(s)**:
	I.15. Description of commodity:		I.16. Commodity code (HS Code): I.17. Total quantity:
	I.18. Temperature of commodities for transport Chilled <input type="checkbox"/> Frozen <input type="checkbox"/>		I.19. Total number of packages:
	I.9. Place of origin: Name: Address:		I.21. Type of packaging:
I.22. Commodities intended for use as: Artificial Reproduction <input type="checkbox"/> Other <input type="checkbox"/>			
I.23. Not Applicable			
I. 24. Identification of commodities: Species (Scientific name) <input type="checkbox"/> <input type="checkbox"/>	Nature of commodity:  Approval number of establishments:	Treatment type:	
Number of packages:	Net weight:	Lot ID/date code:	

Part II: Detail of donors	<b>Donor information</b>																						
	Name		Donor identification		Breed			Date of Birth			Country of Birth			Name of Owner			Address of Owner						
	<b>Semen information</b>																						
	Donor identification		Date/s of collection		Straw/pellet identification			Number of Straws/pellets		Date of entry into semen collection facility			Name of semen collection facility			Address of semen collection facility			Semen collection facility approval number		Date of last inspection of semen facility		
	<b>Test information</b>																						
	(Note that this information is to be amended as appropriate to the exporting country)																						
			<Disease name>			<Disease name>			<Disease name>			<Disease name>			<Disease name>			<Disease name>			<Disease name>		
	Donor identification		Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result
	<b>Other information</b>																						
	<disease name> Vaccine					Scrapie																	
Name of the vaccine		Inactivated or modified live virus		Virus types and strains		Genotype		Identifying laboratory															



<b>Part III: Specific Requirements</b>	<b>Certificate reference number:</b>
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I, ....., a veterinarian authorised by the veterinary authority certify, after due enquiry that the semen described above satisfy(ies) the following requirements:

**Eligibility**

- (1) The semen is from *Ovis aries* and/or *Capra hircus* (delete as appropriate and initial).
- (2) The semen is frozen and non-genetically modified.

**Diagnostic testing, vaccination, and treatment**

- (3) All required laboratory testing was conducted at a laboratory approved to conduct export testing by the Competent Authority of a country approved to export to New Zealand.
- (4) Tests used must be listed in and carried out in accordance with MPI-STD-TVTL.
- (5) Copies of laboratory reports, or an endorsed, tabulated summary, including test date, type, and results for each donor, are attached to this veterinary certificate.
- (6) All products and vaccinations administered to meet specific disease requirements were administered according to the manufacturer's instruction in a country approved to export to New Zealand. Vaccinations were either the final dose of a primary course or the recommended booster to complement the primary.
  - a) Product name, manufacturer, active ingredient (where applicable) \_\_\_\_\_  
Dose and date of treatment \_\_\_\_\_
  - b) Vaccine name and virus type and strain: \_\_\_\_\_  
Inactivated or modified live virus (circle or delete as appropriate and initial)

**Semen collection facility requirements**

- (7) The semen collection facility met the conditions specified in the OIE *Code* Chapter on general hygiene in semen collection and processing centres.
- (8) The semen collection facility was:
  - a) approved for export by the Competent Authority;
  - b) subject to regular inspection by an Official Veterinarian; and
  - c) under the supervision of a semen collection facility veterinarian approved by the Competent Authority.
- (9) The names and approval numbers of these semen collection facilities are recorded in this veterinary certificate.
- (10) When donors were transferred from one approved semen collection facility to another of equal health status without isolation or testing, the following conditions were applied:
  - a) Donors were examined, by the approved semen collection facility veterinarian, and showed no clinical sign of disease on the day of entry into the facility.
  - b) Transfer was direct.
  - c) Transfer was not through a bluetongue infected zone or donors were protected from insect attack during transit.
  - d) Donors were not in direct or indirect contact with animals of a lower health status.
  - e) The means of transport used was disinfected before use.

**Donor and semen collection facility health status**

- (11) Prior to admission to the semen collection facility, the donors were isolated for at least 28 days at a place specifically approved for this purpose by the Competent Authority. During this time they were not used for natural mating and were isolated from animals not of equivalent health status.
- (12) The approved semen collection facility veterinarian ensured that, on the day(s) of collection of the semen, the health status of each donor was monitored and recorded, and the donor did not show any clinical evidence of infectious diseases transmissible in semen.

**Semen collection, processing, storage and transport**

- (13) Semen was collected and processed in accordance with the current recommendations of the OIE *Code*, unless indicated otherwise in this IHS.

- (14) Antibiotics were added to the semen diluent in accordance with the OIE *Code* Chapter on collection and processing of ovine/caprine semen.

Name and concentration of antibiotics:

\_\_\_\_\_  
\_\_\_\_\_

- (15) None of the cryogenic or cooling agents have been previously used in association with any other product of animal origin.
- (16) Dry ice and associated equipment to process semen pellets have been managed to prevent contamination with semen donors not of equivalent tested health status (delete as appropriate and initial).
- (17) Semen is in new or sanitised containers, which are sealed and tamper evident, and clearly and permanently marked to identify the donor and the date(s) of collection. A code is used for this information and its decipher accompanies the consignment (delete as appropriate and initial). The marking is in accordance with the OIE *Code* and conforms to the international standards of the International Committee for Animal Recording (ICAR; [www.icar.org](http://www.icar.org)).
- (18) Semen was only stored with germplasm that was approved for export to New Zealand. Containers were held until export in storage place approved by the Competent Authority of the exporting country.
- (19) Semen was placed in a container which is sanitised and free of contamination.

Disinfectant (active chemical) and date (delete and initial if the container was new):

\_\_\_\_\_

- (20) Semen was transferred from one transport container to another for further processing (delete if semen was not transferred).

Transfer date, facility, and reason:

\_\_\_\_\_

- (21) The transport container in which the semen is transported to New Zealand was sealed by either the semen collection facility veterinarian or an official veterinarian, using tamper-evident seals.

Seal number \_\_\_\_\_

- (22) Germplasm in this consignment originates from a different country than the country of origin (delete as appropriate and initial). The country of origin is currently approved to export to New Zealand and a letter from the Competent Authority or a certified New Zealand export certificate indicating its compliance with New Zealand requirements accompanies the consignment. The germplasm in this consignment has been verified as the same germplasm originating from <insert name of country of origin> and since its arrival it has been stored in the facility stated in Part 1, under the supervision of the Competent Authority.

## **SPECIFIC REQUIREMENTS FOR IDENTIFIED RISK ORGANISMS:**

### **Bluetongue virus (BTV)**

- (23) Donors were:

- resident in a BTV free country or zone in accordance with the requirements of the OIE *Code*, for at least the 60 days prior to and during collection; or
- resident during the seasonally free period in a BTV seasonally free zone in accordance with the requirements of the OIE *Code*, for at least the 60 days prior to collection; or
- resident in a vector-protected facility for at least the 60 days prior to collection and the facility was regularly inspected and certified as being free from *Culicoides* spp. throughout the period when the donors were resident; or
- subjected to a test listed in MPI-STD-TVTL, with negative results.

### **Crimean Congo haemorrhagic fever virus (CCHF)**

- (24) Donors were resident in a country:

- recognised by the Competent Authority as free from CCHF for the 21 days before collection; and
- where CCHF is officially notifiable; or

- (25) Donors were shorn (as required for tick inspection), including the head and lower legs, treated with an acaricide, listed in MPI-STD-TVTL, and systematically inspected, under Official Veterinarian supervision, to ensure they were free from ticks before entering the vector-proof facility, where they were held for at least 21 days before the first collection. The facility was regularly inspected and tick-free throughout the period when the donors were resident.

### **Foot and mouth disease virus (FMD)**

- (26) Donors were resident for the 3 months before semen collection in a country or zone that is free from FMD without vaccination in accordance with the MPI List of FMD Free Countries and donors showed no clinical signs of FMD on the day of semen collection and the following 30 days; or

(27) Donors were resident for the 3 months before semen collection in a country or zone that is free from FMD with vaccination in accordance with the MPI List of FMD-Free Countries and neither the donors nor any other animal in the collection facility:

- a) showed clinical signs of FMD on the day of the semen collection for New Zealand and for the following 30 days; and
- b) was vaccinated within the month prior to semen collection for New Zealand; and donors have either
  - i) never been vaccinated and were subjected, not less than 21 days after semen collection, to a test for antibodies against FMD virus described in MPI-STD-TVTL; or
  - ii) been vaccinated at least twice, with the most recent vaccination not more than 12 months before collection for New Zealand.

#### **Jaagsiekte sheep retrovirus (ovine pulmonary adenomatosis)**

(28) Donors were resident since birth in countries recognised by the Competent Authority as free from jaagsiekte; or

(29) Donors were resident:

- a) in a country where jaagsiekte is notifiable;
- b) only in premises that have remained free from jaagsiekte for at least the 10 years prior to collection and no sheep/goat from a flock/herd of inferior health status was introduced during that period; and
- c) in premises that include animals over 5 years of age; or

(30) Donors were subjected to a jaagsiekte test listed in MPI-STD-TVTL, with negative results.

#### **Maedi-visna virus (MV)**

(31) Donors were resident since birth in countries recognised by the Competent Authority as free from MV; or

(32) Donors:

- a) only resided with herds/flocks, during the 3 years before collection for New Zealand, where MV was neither clinically nor serologically diagnosed and animals of inferior health status were not introduced;
- b) over one year of age were subject to a MV test listed in MPI-STD-TVTL, with negative results, during the 30 days prior to entering the isolation facility; and
- c) were subjected to a MV test listed in MPI-STD-TVTL, with negative results, at least 21 days after entering isolation and at least annually thereafter while in the collection facility.

#### **Peste des petits ruminants virus (PPR)**

(33) Donors were resident in a PPR free country or zone in accordance with the OIE *Code* for at least 21 days prior to and during collection; or

(34) Donors:

- a) were resident in an establishment not located in a PPR infected zone in accordance with the OIE *Code*; and
- b) showed no clinical signs of PPR on the day of collection and during the following 21 days and during that period no case of PPR was officially reported in that establishment; and donors were either:
  - i) vaccinated against PPR between 21 and 120 days prior to semen collection; or
  - ii) unvaccinated and subjected to a test listed in MPI-STD-TVTL at least 21 days after semen collection, with negative results

#### **Rift Valley fever virus (RVF)**

(35) Donors were resident in a RVF free country or zone in accordance with the OIE *Code* for at least the 30 days prior to collection; or

(36) Donors were held in a vector-proof facility for at least 30 days prior to and during collection and never showed clinical signs of RVF. The facility was inspected regularly and mosquito-free throughout the period when donors were resident; or

(37) For at least the 28 days prior to and after semen collection, the donors showed no clinical sign of RVF; and either

- a) Donors were serologically tested for RVF, using a test listed in MPI-STD-TVTL, on the day of semen collection, and at least 14 days later, and showed no significant rise in titre; or
- b) Donors were vaccinated against RVF in accordance with the OIE *Manual*, at least 21 days prior to semen collection with a modified live vaccine.

#### **Capripox virus (sheep and goat pox)**

(38) Donors were resident in a sheep and goat pox free country in accordance with the OIE *Code* for at least the 21 days prior to collection; or

(39) Donors showed no clinical signs of sheep or goat pox on the day of collection of the semen and for the following 21 days; and

- a) For at least the 21 days prior to collection, the donor:
  - i) resided in an establishment where no case of sheep or goat pox was reported during that period;
  - ii) was not in a zone infected with sheep and goat pox in accordance with the OIE *Code*; and

- b) vaccinated donors were vaccinated in accordance with the OIE *Manual*.

#### **Vesicular stomatitis virus (VS)**

- (40) Donors were resident in a country or zone that is free from VS in accordance with the OIE *Code* for at least the 21 days prior to collection; or  
(41) Donors were resident in a flock/herd where no case of VS was reported at the time of collection and for the 30 days after collection.

#### **Wesselsbron virus**

- (42) Donors were resident in a country recognised by the Competent Authority as free from circulating Wesselsbron disease virus (serological surveys required) for at least the 21 days prior to collection; or  
(43) Donors were resident in an establishment where Wesselsbron disease has not occurred for at least the 21 days prior to collection.

#### ***Brucella melitensis* (caprine and ovine brucellosis)**

- (44) Donors were resident in a country, zone, or flock/herd that is officially free from caprine and ovine brucellosis in accordance with the OIE *Code*; or  
(45) Donors were resident in a flock/herd that is free from caprine and ovine brucellosis, in accordance with the OIE *Code*, and were subjected to two different tests for caprine and ovine brucellosis listed in MPI-STD-TVTL on the same blood sample, taken within 30 days prior to semen collection, with negative results.

#### ***Mycoplasma capricolum subsp. Capripneumoniae* (contagious caprine pleuropneumonia - CCPP)**

- (46) For goats only:
- a) Donors were resident in a country that is free from CCPP in accordance with the OIE *Code*; or
  - b) For at least the 45 days prior to collection, donors did not reside in a CCPP infected zone, in accordance with the OIE *Code*, and were not resident in a herd where CCPP had been officially reported during that time; and
    - i) Aliquots of semen from each collection were subjected to a test listed in MPI-STD-TVTL, with negative results; or
    - ii) Donors were subjected to a CCPP complement fixation test, in accordance with the OIE *Manual*, on two occasions, with an interval of 21 to 30 days between tests and the second test being within the 14 days prior to pre-entry isolation, with negative results; and
      - i. Donors were isolated from other domestic goats from the first test until the last date of collection.

#### ***Mycoplasma agalactiae* (contagious agalactia)**

- (47) Donors were resident in a country that has been recognised by the Competent Authority as free from contagious agalactia for at least the 6 months prior to collection; or  
(48) Donors were:
  - a) resident for at least the 6 months prior to collection only at premises where no case of contagious agalactia had been officially reported during that time; and
  - b) tested for *Mycoplasma agalactiae* using a test listed in MPI-STD-TVTL.

#### ***Mycobacterium caprae* and *Mycobacterium bovis* (tuberculosis)**

- (49) For goats only:
- a) Donors were resident in a country recognised by the Competent Authority as free from tuberculosis in goats; or
  - b) Donors were subjected to a single comparative tuberculin test for tuberculosis prior to entry to the collection facility, with negative results; and
    - i) All animals in the collection facility were tested prior to entry and at least annually, with negative results.

#### ***Chlamydia abortus* (enzootic abortion of ewes - EAE)**

- (50) Donors were resident in a country recognised by the Competent Authority as free from EAE for at least the 2 years prior to collection; or  
(51) Donors were:
  - a) resident in a herd/flock that is free from EAE in accordance with the OIE *Code* for at least the past 2 years;
  - b) not in contact with any animal of lower health status during that period of time; and either
    - i) subjected to a test for EAE listed in MPI-STD-TVTL, with negative results; or
    - ii) Semen samples for each collection were subjected to a test for EAE listed in MPI-STD-TVTL, with negative results.

#### ***Coxiella burnetii* (Q fever)**

(52) Donors have never been confirmed positive for Q fever; and either

- a) Donors were subjected to a Q fever listed in MPI-STD-TVTL, with negative results; or
- b) Semen from each collection was subjected to a Q fever test listed in MPI-STD-TVTL, with negative results.

### Scrapie

(53) For goats only:

- a) Donors were resident in a scrapie free country in accordance with the OIE Code; or
- b) Donors were resident in an establishment that has been maintained free from scrapie from commencement until conclusion of collection, in accordance with the OIE Code recommendations for a scrapie free establishment.

(54) For sheep only:

- a) Donors were resident in a scrapie free country in accordance with the OIE Code; or
- b) Donors were resident in an establishment that has been maintained free from scrapie from commencement until conclusion of collection, in accordance with the OIE Code recommendations for a scrapie free establishment; or
- c) Donors have the scrapie resistant genotypes – ARR/ARR, ARR/AHQ, ARR/ARH or ARR/ARQ. Laboratory evidence of the genotype is required.

### Semen Facility Veterinarian:

Name

Address (in capital letters):

Date:

Signature:

### Official Veterinarian:

Name and address (in capital letters):

Date:

Signature:

Official Veterinarian signature

## 5.8 Model Veterinary Certificate for International Trade in Embryos from Sheep (*Ovis aries*) and Goats (*Capra hircus*)

- (1) Below is the model veterinary certificate for trade in embryos from sheep (*Ovis aries*) and goats (*Capra hircus*). The model meets the requirements of the IHS.
- (2) The model veterinary certificate format is based on the OIE *Code* Chapter for model veterinary certificates for international trade in germplasm.

COUNTRIES:			
Part I: Details of dispatched consignment	I.1. Consignor (Exporter): Name: Address:		I.2. Certificate reference number:
			I.3. Competent Authority:
	I.4. Consignee (Importer): Name: Address:		
	I.5. Country of origin ISO Code*	I.6. Zone or compartment of origin**:	
	I.7. Country of destination: ISO Code*	I.8. Zone or compartment of destination**:	
	I.9. Place of origin: Name: Address:		
	I.10. Place of shipment:	I.11. Date of departure:	
	I.12. Means of transport: Aeroplane <input type="checkbox"/> Ship <input type="checkbox"/> Identification:	I.13. Expected border post:	
			I.14. CITES permit No(s)**:
	I.15. Description of commodity:	I.16. Commodity code (HS Code):	
		I.17. Total quantity:	
I.18. Temperature of commodities for transport Chilled <input type="checkbox"/> Frozen <input type="checkbox"/>	I.19. Total number of packages:		
I.9. Place of origin: Name: Address:	I.21. Type of packaging:		
I.22. Commodities intended for use as: Artificial Reproduction <input type="checkbox"/> Other <input type="checkbox"/>			
I.23. Not Applicable			
I. 24. Identification of commodities: Species (Scientific name) <input type="checkbox"/> <input type="checkbox"/>	Nature of commodity:  Approval number of establishments:	Treatment type:	
Number of packages:	Net weight:	Lot ID/date code:	

## Part II: Details of donors

Female donor information																						
Name	Donor identification			Breed			Date of Birth			Country of Birth			Name of Owner			Address of Owner						
Male donor information																						
Name	Donor identification			Breed			Date of Birth			Country of Birth			Name of Semen Facility			Address of Semen Facility			Semen Facility Number			
Embryo information																						
Female donor identification	Date/s of collection			Straw identification			Number of Straws			Number of Embryos /Straws			Name and Address of Embryo Collection Flock/Herd			Male donor identification			Date of Semen Collection or date of Natural Mating			
Test information																						
(Note that this information is to be amended as appropriate to the exporting country)																						
		<Disease name>			<Disease name>			<Disease name>			<Disease name>			<Disease name>			<Disease name>			<Disease name>		
Donor Identification	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	
Other information																						
<disease name> Vaccine					Scrapie																	
Name of the vaccine	Inactivated or modified live virus	Virus types and strains	Genotype	Identifying laboratory																		

<b>Part III: Specific Requirements</b>	<b>Certificate reference number:</b>
<p>I,....., a veterinarian authorised by the veterinary authority certify, after due enquiry that the semen described above satisfy(ies) the following requirements:</p>	
<p><b>Eligibility</b></p> <p>(1) The embryos are from <i>Ovis aries</i> or <i>Capra hircus</i> (delete as appropriate and initial).</p> <p>(2) The embryos are <i>in vivo</i> derived, frozen, non-cloned, and non-genetically modified.</p>	
<p><b>Diagnostic testing, vaccination, and treatment</b></p> <p>(3) All required laboratory testing was conducted at a laboratory approved to conduct export testing by the Competent Authority of a country approved to export to New Zealand.</p> <p>(4) Tests used must be listed in and carried out in accordance with MPI-STD-TVTL.</p> <p>(5) Copies of laboratory reports, or an endorsed, tabulated summary, including test date, type, and results for each donor, are attached to this veterinary certificate.</p> <p>(6) All products and vaccinations administered to meet specific disease requirements were administered according to the manufacturer's instruction in a country approved to export to New Zealand. Vaccinations were either the final dose of a primary course or the recommended booster to complement the primary.</p> <p style="margin-left: 40px;">a) Product name, manufacturer, active ingredient (where applicable) _____</p> <p style="margin-left: 80px;">Dose and date of treatment _____</p> <p style="margin-left: 40px;">b) Vaccine name and virus type and strain: _____</p> <p style="margin-left: 80px;">Inactivated or modified live virus (circle or delete as appropriate and initial)</p>	
<p><b>Embryo collection team and herd approval requirements</b></p> <p>(7) At the time of collection of embryos for export to New Zealand, the embryo collection team was approved by and registered with the Competent Authority of the exporting country.</p> <p>(8) The Competent Authority has knowledge of and authority over the embryo collection flock/herd until completion of collection and testing specified in this IHS.</p>	
<p><b>Donor and flock/herd health status</b></p> <p>(9) Donors were resident in the embryo collection flock/herd for at least 28 days prior to collection of embryos for export to New Zealand.</p> <p>(10) Where a specific requirement for a risk organism is met by pre-collection testing, donors were isolated from other sheep or goats not of an equivalent tested health status, from the time of the pre-collection test until completion of collection of embryos for export to New Zealand.</p> <p>(11) On the day(s) of collection of the embryos, the approved embryo collection team veterinarian was responsible for monitoring the health status of each donor and recording that the donor was free from clinical evidence of infectious diseases transmissible in embryos.</p> <p>(12) The semen used to produce the embryos in the consignment either:</p> <p style="margin-left: 40px;">a) was imported directly from New Zealand or is eligible for export to New Zealand; or</p> <p style="margin-left: 40px;">b) was collected, processed, and stored at a semen collection facility that complies with the official semen collection facility protocols of the exporting country (where MPI deems this to be equivalent); or</p> <p style="margin-left: 40px;">c) Where natural service or fresh semen was used, donor males were inspected, found free from clinical evidence of infectious diseases transmissible in semen, and satisfy the testing and isolation requirements for semen from sheep and goats.</p>	
<p><b>Embryo collection, processing, storage and transport</b></p> <p>(13) Embryos were collected and processed under the supervision of an approved embryo collection team veterinarian and in accordance with the recommendations in the OIE <i>Code</i> chapters on collection and processing of <i>in vivo</i> derived embryos of livestock.</p> <p>(14) Embryos were treated with trypsin during the washing process as described in the IETS <i>Manual</i>.</p> <p>(15) Embryos had an intact zona pellucida and were free of adherent material after the final wash when examined over its entire surface at not less than 50X magnification. Any micro-manipulation that caused a breach of the zona pellucida, was performed according to the procedures described in the OIE <i>Code</i> and IETS <i>Manual</i>.</p> <p>(16) All biological products of animal origin used in the media and solutions for collection, processing, washing or storage of embryos was free from pathogenic organisms including pestiviruses.</p>	



- (17) Media and solutions were either sterilised by approved methods according to the IETS *Manual* or commercially prepared sterile media were used. These were handled in such a manner as to ensure that sterility was maintained.
- (18) Antibiotics as recommended in the OIE *Code* and IETS *Manual*, or a combination of antibiotics with equivalent activity, were added to collection, processing, washing and storage media.
- Name and concentration of antibiotics:
- \_\_\_\_\_
- \_\_\_\_\_
- (19) None of the cryogenic or cooling agent has been previously used in association with any other product of animal origin.
- (20) Embryos are sealed in straws, which are clearly and permanently marked to identify the donor and the date(s) of collection. A code is used for this information and its decipher accompanies the consignment (delete as appropriate and initial). The marking is in accordance with the OIE *Code* and conforms to the international standards of the International Committee for Animal Recording (ICAR; [www.icar.org](http://www.icar.org)).
- (21) Embryos were only stored with germplasm that was approved for export to New Zealand. Containers were held until export in a storage place approved by the Competent Authority of the exporting country.
- (22) Embryos were transferred from one transport container to another for further processing (delete if embryos were not transferred).
- Transfer date, location, and reason:
- \_\_\_\_\_
- (23) Embryos were placed in a container which is sanitised and free of contamination.
- Disinfectant (active chemical) and date (delete and initial if container was new):
- \_\_\_\_\_
- (24) The transport container in which the embryos are transported to New Zealand was sealed by either the embryo collection team veterinarian or an official veterinarian, using tamper evident seals.
- Seal number \_\_\_\_\_
- (25) Germplasm in this consignment originates from a different country than the country of origin (delete as appropriate and initial). The country of origin is currently approved to export to New Zealand and a letter from the Competent Authority or a certified New Zealand export certificate indicating its compliance with New Zealand requirements accompanies the consignment. The germplasm in this consignment has been verified as the same germplasm originating from <insert name of country of origin> and since its arrival it has been stored in the facility stated in Part 1, under the supervision of the Competent Authority.

#### **SPECIFIC REQUIREMENTS FOR IDENTIFIED RISK ORGANISMS:**

##### **Bluetongue virus (BTV)**

- (26) Donors were:
- resident in a BTV free country or zone in accordance with the requirements of the OIE *Code*, for at least the 60 days prior to and during collection; or
  - resident during the seasonally free period in a BTV seasonally free zone in accordance with the requirements of the OIE *Code*, for at least the 60 days prior to collection; or
  - resident in a vector-protected facility for at least the 60 days prior to collection and the facility was regularly inspected and certified as being free from *Culicoides* spp. throughout the period when the donors were resident; or
  - subjected to a test listed in MPI-STD-TVTL, with negative results.

##### **Crimean Congo haemorrhagic fever virus (CCHF)**

- (27) Donors were resident in a country:
- recognised by the Competent Authority as free from CCHF for the 21 days before collection; and
  - where CCHF is officially notifiable; or
- (28) Donors were shorn (as required for tick inspection), including the head and lower legs, treated with an acaricide, listed in MPI-STD-TVTL, and systematically inspected, under Official Veterinarian supervision, to ensure they were free from ticks before entering the vector-proof facility, where they were held for at least 21 days before the first collection. The facility was regularly inspected and tick-free throughout the period when the donors were resident.

##### **Foot and mouth disease virus (FMD)**

- (29) Donors were resident for the 3 months before embryo collection in a country or zone that is free from FMD without vaccination in accordance with the MPI List of FMD-Free Countries and Zones and donors showed no clinical signs of FMD on the day of embryo

collection and the following 30 days; or

- (30) Donors were resident for the 3 months before embryo collection in a country or zone that is free from FMD with vaccination in accordance with the MPI List of FMD-Free Countries and neither the donors nor any other animal at the collection herd/flock:
- a) showed clinical signs of FMD on the day of the embryo collection for New Zealand and for the following 30 days; and
  - b) was vaccinated within the month prior to embryo collection for New Zealand; and donors have either
    - i) never been vaccinated and were subjected, not less than 21 days after embryo collection, to a test for antibodies against FMD virus described in MPI-STD-TVTL; or
    - ii) been vaccinated at least twice, with the most recent vaccination not more than 12 months before collection for New Zealand.

#### **Jaagsiekte sheep retrovirus (ovine pulmonary adenomatosis)**

- (31) Donors were resident since birth in countries recognised by the Competent Authority as free from jaagsiekte; or
- (32) Donors were resident:
- a) in a country where jaagsiekte is notifiable;
  - b) only in premises that have remained free from jaagsiekte for at least the 10 years prior to collection and no sheep/goat from a flock/herd of inferior health status was introduced during that period; and
  - c) in premises that include animals over 5 years of age; or
- (33) Donors were subjected to jaagsiekte test listed in MPI-STD-TVTL, with negative results.

#### **Maedi-visna virus**

- (34) Donors were resident since birth in countries recognised by the Competent Authority as free from MV; or
- (35) Donors:
- a) only resided with herds/flocks, during the 3 years before collection for New Zealand, where MV was neither clinically nor serologically diagnosed and animals of inferior health status were not introduced;
  - b) over one year of age were subjected to a MV test listed in MPI-STD-TVTL, with negative results, during the 30 days prior to entering the collection herd/flock; and
  - c) were subjected to a MV test listed in MPI-STD-TVTL, with negative results, at least 21 days after entering the collection herd/flock and at least annually thereafter while in the collection herd/flock.

#### **Peste des petits ruminants virus (PPR)**

- (36) Donors were resident in a PPR free country or zone in accordance with the OIE Code for at least 21 days prior to and during embryo collection; or
- (37) Donors:
- a) were resident in an establishment not located in a PPR infected zone in accordance with the OIE Code;
  - b) showed no clinical signs of PPR on the day of embryo collection and during the following 21 days and during that period no case of PPR was officially reported in that establishment; and donors were either
    - i) vaccinated against PPR between 21 and 120 days prior to embryo collection; or
    - ii) unvaccinated and subjected to a test listed in MPI-STD-TVTL at least 21 days after embryo collection, with negative results.

#### **Rift Valley fever virus (RVF)**

- (38) Donors were resident for at least the 30 days prior to embryo collection in a country or zone that is free from RVF in accordance with the OIE Code; or
- (39) Donors were held in an MPI approved vector-proof collection facility for at least 30 days prior to and during collection and never showed clinical signs of RVF. The facility was inspected regularly and mosquito-free throughout the period when donors were resident.
- (40) For at least the 28 days prior to and after embryo collection, the donors showed no clinical sign of RVF; and either
- a) Donors were serologically tested for RVF, using a test listed in MPI-STD-TVTL, on the day of embryo collection, and at least 14 days later, and showed no significant rise in titre; or
  - b) Donors were vaccinated against RVF in accordance with the OIE Manual, at least 21 days prior to embryo collection with a modified live vaccine.

#### **Capripox virus (sheep and goat pox)**

- (41) Donors were resident in a sheep and goat pox free country in accordance with the OIE Code for at least the 21 days prior to collection; or
- (42) Donors showed no clinical signs of sheep or goat pox on the day of embryo collection and for the following 21 days; and
- a) For at least the 21 days prior to collection, the donor:

- i) resided in an establishment where no case of sheep or goat pox was reported during that period;
  - ii) was not in a zone infected with sheep and goat pox in accordance with the OIE Code; and
- b) vaccinated donors were vaccinated in accordance with the OIE Manual.

#### **Vesicular stomatitis virus (VS)**

- (43) Donors were resident in a country that is free from VS in accordance with the OIE Code for at least the 21 days prior to collection; or
- (44) Donors were resident in a flock/herd where no case of VS was reported at the time of collection and for the 30 days after collection.

#### **Wesselsbron virus**

- (45) Donors were resident in a country recognised by the Competent Authority as free from Wesselsbron disease virus (serological surveys required) for at least the 21 days prior to collection; or
- (46) Donors were resident in an establishment where Wesselsbron disease has not occurred for at least the 21 days prior to collection.

#### **Brucella melitensis (caprine and ovine brucellosis)**

- (47) Donors were resident in a country, zone, or flock/herd that is officially free from caprine and ovine brucellosis in accordance with the OIE Code; or
- (48) Donors were resident in a flock/herd that is free from caprine and ovine brucellosis, in accordance with the OIE Code, and were subjected to two different tests for caprine and ovine brucellosis listed in MPI-STD-TVTL on the same blood sample, taken within 30 days prior to embryo collection, with negative results.

#### **Mycoplasma capricolum subsp. Capripneumoniae (contagious caprine pleuropneumonia - CCPP)**

- (49) For goats only:
- a) Donors were resident in a country that is free from CCPP in accordance with the OIE Code; or
  - b) For at least the 45 days prior to collection, donors did not reside in a CCPP infected zone, in accordance with the OIE Code, and were not resident in a herd where CCPP had been officially reported during that time; and
    - i) Aliquots of embryos/oocytes or collection/washing fluids from each collection were subjected to a test listed in MPI-STD-TVTL, with negative results; or
    - ii) Donors were subjected to a CCPP complement fixation test, in accordance with the OIE Manual, on two occasions, with an interval of 21 to 30 days between tests and the second test being within the 14 days prior to pre-entry isolation, with negative results; and
      - i. Donors were isolated from other domestic goats from the first test until the last date of collection.

#### **Mycoplasma agalactiae (contagious agalactia)**

- (50) Donors were resident in a country that has been recognised by the Competent Authority as free from contagious agalactia for at least the 6 months prior to collection; or
- (51) Donors were:
- a) resident for at least the 6 months prior to collection only at premises where no case of contagious agalactia had been officially reported during that time; and
  - b) tested for Mycoplasma agalactiae using a test listed in MPI-STD-TVTL.

#### **Mycobacterium caprae and Mycobacterium bovis (tuberculosis)**

- (52) For goats only:
- a) Donors were resident in a country recognised by MPI as being free from tuberculosis in goats; or
  - b) Donors were subjected to a single comparative tuberculin test for tuberculosis prior to entry to the collection flock/herd, with negative results; and
    - i) All animals in the embryo collection flock/herd were tested prior to entry and at least annually, with negative results.

#### **Chlamydia abortus (enzootic abortion of ewes - EAE)**

- (53) Donors were resident in a country recognised by the Competent Authority as free from EAE for at least the 2 years prior to collection; or
- (54) Donors were:
- a) resident in a herd/flock that is free from EAE in accordance with the OIE Code for at least the past 2 years.
  - b) not in contact with any animal of lower health status during that period of time; and either

- i) subjected to a test for EAE listed in MPI-STD-TVTL, with negative results; or
- ii) Samples of embryos/oocytes or collection/washing fluids from each embryo collection were subjected to a test for EAE listed in MPI-STD-TVTL, with negative results.

***Coxiella burnetii* (Q fever)**

(55) Donors have never been confirmed positive for Q fever; and either

- a) Donors were subjected to a Q fever test listed in MPI-STD-TVTL, with negative results; or
- b) Embryos/oocytes or collection/washing fluids from each collection were subjected to a Q fever test listed in MPI-STD-TVTL, with negative results.

**Scrapie**

(56) For goats only:

- a) Donors were resident in a country that is free from scrapie in accordance with the OIE *Code*; or
- b) Donors were resident in a collection herd/flock that has been maintained free from scrapie from commencement until conclusion of collection, through compliance with the OIE *Code* recommendations for a scrapie free establishment; or
- c) Embryos comply with the OIE *Code* recommendations for importation of *in vivo* derived goat embryos from countries or zones not considered free from scrapie.

**Embryo Collection Veterinarian:**

Name

Address (in capital letters):

Date:

Signature:

**Official Veterinarian:**

Name and address (in capital letters):

Date:

Signature:



## Appendix 1 – Change History

Previous Version Date	Current Version Date	Section Changed	Change(s) Description

Draft for Consultation